

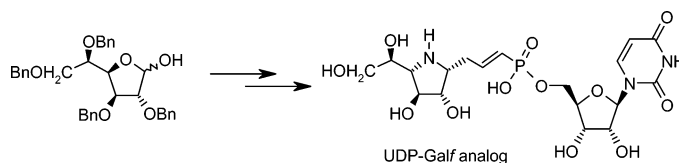
Convergent and Stereoselective Synthesis of Iminosugar-Containing Galf and UDP-Galf Mimicks: Evaluation as Inhibitors of UDP-Gal Mutase

Virginie Liautard,[†] Valérie Desvergnès,^{*,†,‡} Kenji Itoh,[§] Hung-wen Liu,[§] and Olivier R. Martin^{*,†}

Institut de Chimie Organique et Analytique, Université d'Orléans, CNRS-UMR 6005, BP 6759, 45067 Orléans, France, and Division of Medicinal Chemistry, College of Pharmacy and Department of Chemistry and Biochemistry, University of Texas at Austin, Austin, Texas 78712

olivier.martin@univ-orleans.fr; v.desvergnès@ism.u-bordeaux1.fr

Received January 17, 2008



The synthesis of a UDP-Galf analog incorporating a 1,4-dideoxy-1,4-imino-D-galactitol skeleton α -linked to UMP by a 3C-tether and of a series of related pyrrolidine galactofuranose mimicks is reported. These compounds were obtained by way of the highly stereoselective reaction of silylated nucleophiles with a *N*-Cbz glucufuranosylamine which afforded the corresponding open-chain product with a 1,2-*syn* stereochemistry, as predicted from pioneering studies from Kobayashi. Cyclization of these intermediates afforded α -C-glycosides of imino-galactofuranose carrying various functional groups in the aglycone. Further elaboration of the α -C-allyl substituted derivative by cross-metathesis with a uridin-5'-yl vinylphosphonate provided, after deprotection, the desired original UDP-Galf mimicks. Cleavage of the benzyl ether protecting groups in the iminosugar component using BCl_3 proved critical to the success of the synthetic plan. Several of the new 1,4-dideoxy-1,4-imino-D-galactitol derivatives were evaluated as inhibitors of UGM (UDP-galactopyranose mutase) from *Escherichia coli*; however, none of them exhibited less than mM activities toward this enzyme which catalyzes a crucial step of the biosynthesis of galactofuranose-containing bacterial cell-surface glycans.

Introduction

The mycobacterial cell wall exhibits a lipidated polysaccharidic structure unique in the prokaryote reign that is essential for the microorganism growth and survival.¹ This mycolyl-arabinogalactan-peptidoglycan (mAGP) complex includes a galactan biopolymer composed of D-galactofuranose units (Galf), a ring-form of galactose that occurs much less frequently than its pyranose form (Galp). Members of the genus *Mycobacterium* are responsible for major health problems such as tuberculosis

and leprosy. In the context of the fight against such diseases,² which is becoming even more critical as a result of the appearance of drug-resistant strains,³ inhibition of the biosynthetic pathway leading to galactofuranose-containing glycans has been identified as a promising new therapeutic approach.⁴ Indeed, the key intermediate of this pathway, UDP-galactofuranose (Scheme 1), is absent from mammals;⁵ it is formed from UDP-galactopyranose by a ring contraction process catalyzed by UDP-galactopyranose mutase (UPM)⁶ and in the next step,

* To whom correspondence should be addressed. Tel: ++ 33 2 38 49 45 81. Fax: ++ 33 2 38 41 72 81 (O.R.M.); Tel: ++ 33 5 40 00 62 87 (V.D.).
[†] Université d'Orléans.

[‡] Present address: Institut des Sciences Moléculaires, Université de Bordeaux 1, CNRS-UMR 5255, 351 cours de la libération, 33405 Talence, France.

[§] University of Texas at Austin.

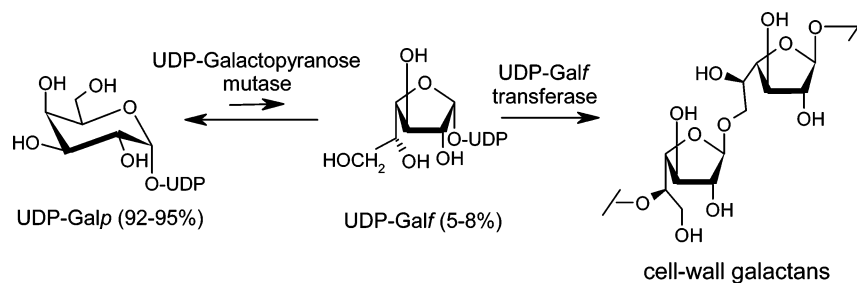
(1) (a) Rastogi, N.; Goh, K. S.; David, H. L. *Antimicrob. Agents Chemother.* **1990**, *34*, 759–764. (b) Barry, C. E. *Biochem. Pharmacol.* **1997**, *54*, 1165–1172. (c) Pan, F.; Jackson, M.; Ma, Y.; McNeil, M. *J. Bacteriol.* **2001**, *183*, 3991–3998. (d) Brennan, P. J. *Tuberculosis* **2003**, *83*, 91–97.

(2) (a) O'Brien, R. J.; Nunn, P. P. *Am. J. Respir. Crit. Care Med.* **2001**, *163*, 1055–1058. (b) Zhang, Y.; Amzel, L. M. *Curr. Drug Targets* **2002**, *3*, 131–154. (c) Tripathi, R. P.; Tewari, N.; Dwivedi, N.; Tiwari, V. K. *Med. Res. Rev.* **2005**, *25*, 93–131. (d) Agrawal, Y. K.; Bhatt, H. G.; Raval, H. G.; Oza, P. M.; Vaidya, H. B.; Manna, K.; Gogoi, P. *J. Sci. Ind. Res.* **2007**, *66*, 191–208.

(3) See in particular the recent article in *C&EN* **2007**, *39*, 21–32.

(4) (a) Pedersen, L. L.; Turco, S. J. *Cell. Mol. Life Sci.* **2003**, *60*, 259–266. (b) Brennan, P. J.; Crick, D. C. *Curr. Top. Med. Chem.* **2007**, *7*, 475–488.

SCHEME 1. UDP-Galactofuranose and Galactan Biosynthesis



Galf residues are incorporated into the galactan structure via alternating β -(1 \rightarrow 5) and β -(1 \rightarrow 6) linkage by a bifunctional UDP-Galf transferase.⁷

Despite elegant bioorganic studies,⁸ the mechanism of the unprecedented ring contraction promoted by the mutase has not yet been fully elucidated. According to most recent studies, it appears that the reaction involves a displacement mechanism in which a reduced flavin cofactor acts as a nucleophile, thus substituting UDP from UDP-Galp and leading to a *N*-galactosylated intermediate which allows equilibration between the furanose and pyranose forms of galactose. As in other reactions at the anomeric carbon of glycosyl donors, these displacements are thought to occur by way of an oxocarbenium-like transition state.⁹ Since iminosugars are known to be mimicks of such positively charged states, we considered that pyrrolidine-containing analogs of UDP-Galf might be of interest as potential inhibitors of both the ring contraction and the glycosyl transfer reactions. In support of this proposal, pyrrolidine-based nucleoside analogs have been conceived by Schramm and Tyler¹⁰ as inhibitors of purine nucleoside phosphorylases and related enzymes on the basis of a detailed study of transition state structures carrying a partial positive charge at the reaction site; some of these compounds turned out to be the most powerful

enzyme inhibitors reported to date. More largely, iminosugars behave as inhibitors of a wide range of enzymatic processes and are under investigations as potential therapeutic agents for a diversity of diseases.¹¹

In the context of our studies on iminosugars of biological interest,¹² we engaged in a research program on novel UDP-Galf mimicks containing a 1,4-dideoxy-1,4-imino-D-galactitol entity. While syntheses of the parent 1,4-dideoxy-1,4-imino-D-galactitol and of a homo-derivative have been reported,¹³ a single example of such *galacto*-pyrrolidine linked to uridine has been described so far.¹⁴ We reported recently the first synthesis of a β -linked UDP-Galf mimicks based on a pyrrolidinol skeleton by way of a 1,3-dipolar cycloaddition process onto a cyclic nitrone.¹⁵ We describe in this article the full details of a synthetic study of UDP-Galf mimicks in which a UMP fragment is α -linked to 1,4-dideoxy-1,4-imino-D-galactitol by a 3-carbon tether.¹⁶ The final, fully deprotected UDP-Galf analogs as well as related compounds were then tested as inhibitors of UDP-Gal mutase.

Synthetic Design. Our retrosynthetic analysis starts with D-glucose and is based on reactions of the Z-protected glucosylamine **A** as key intermediate (Scheme 2). The crucial step of the strategy is the highly stereoselective Lewis-acid-catalyzed nucleophilic addition of silylated nucleophiles onto the corresponding *N*-acyl iminium ion, a process pioneered by Kobayashi et al.¹⁷ This process would thus lead to precursors of ' α -configured' 1,4-iminogalactitol derivatives. Then, coupling with UMP would be achieved by cross-metathesis between an α -1-C-allyl-1,4-imino-D-galactitol **B** and a uridin-5'-yl vinylphosphonate to provide eventually compounds of type **C**.

(5) Galf is found in: Trypanosomatids: De Lederkremer, R. M.; Colli, W. *Glycobiology* **1995**, *5*, 547–552; *S. disenteriae*: Dmitriev, B. A.; Lvov, V. L.; Kochetkov, N. K. *Carbohydr. Res.* **1977**, *56*, 207–209; *M. tuberculosis*: Besra, G. S.; Khoo, K. H.; McNeil, M. R.; Dell, A.; Morris, H. R.; Brennan, P. J. *Biochemistry* **1995**, *34*, 4257–4266. Galf has also been found recently in lower eukaryotes: (a) Bakker, H.; Kleczka, B.; Gerardy-Schahn, R.; Routier, F. H. *Biol. Chem.* **2005**, *7*, 657–661. (b) Beverley, S. M.; Owens, K. L.; Showalter, M.; Griffith, C. L.; Doering, T. L.; Jones, V. C.; McNeil, M. R. *Eukaryotic Cell* **2005**, *6*, 1147–1154.

(6) (a) Sanders, D. A. R.; Staines, A. G.; McMahon, S. A.; McNeil, M. R.; Whitfield, C.; Naismith, J. H. *Nat. Struct. Biol.* **2001**, *8*, 858–863. (b) Beis, K.; Srikannathasan, V.; Liu, H.; Fullerton, S. W.; Bamford, V. A.; Sanders, D. A.; Whitfield, C.; McNeil, M. R.; Naismith, J. H. *J. Mol. Biol.* **2005**, *4*, 971–982.

(7) (a) Belanova, M.; Dianiskova, P.; Brennan, P. J.; Completo, G. C.; Rose, N. L.; Lowary, T. L.; Mikusova, K. *J. Bacteriol.* **2008**, *190*, 1141–1145. (b) Rose, N. L.; Completo, G. C.; Lin, S.-J.; McNeil, M.; Palcic, M. M.; Lowary, T. L. *J. Am. Chem. Soc.* **2006**, *128*, 6721–6729. (c) Mikusova, K.; Belanova, M.; Kordulakova, J.; Honda, K.; McNeil, M. R.; Mahapatra, S.; Crick, D. C.; Brennan, P. J. *J. Bacteriol.* **2006**, *188*, 6592–6598. (d) Mikusova, K.; Yagi, T.; Stern, R.; McNeil, M. R.; Besra, G. S.; Crick, D. C.; Brennan, P. J. *J. Biol. Chem.* **2000**, *275*, 33890–33897. (e) Kremer, L.; Dover, L. G.; Morehouse, C.; Hitchin, P.; Everett, M.; Morris, H. R.; Dell, A.; Brennan, P. J.; McNeil, M. R.; Flaherty, C.; Duncan, K.; Besra, G. S. *J. Biol. Chem.* **2001**, *276*, 26430–26440.

(8) (a) Huang, Z.; Zhang, Q.; Liu, H. W. *Bioorg. Chem.* **2003**, *31*, 494–502. (b) Soltero-Higgin, M.; Carlson, E. E.; Gruber, T. D.; Kiessling, L. L. *Nat. Struct. Mol. Biol.* **2004**, *11*, 539–543. (c) Caravano, A.; Sinaÿ, P.; Vincent, S. P. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 1123–1125. (d) Mansoorabadi, S. O.; Thibodeaux, C. J.; Liu, H. W. *J. Org. Chem.* **2007**, *72*, 6329–6342. (e) Chad, J. M.; Sarathy, K. P.; Gruber, T. D.; Addala, E.; Kiessling, L. L.; Sanders, D. A. *Biochemistry* **2007**, *46*, 6723–6732.

(9) Itoh, K.; Huang, Z.; Liu, H. W. *Org. Lett.* **2007**, *9*, 879–882.

(10) Schramm, V. L.; Tyler, P. C. *Curr. Top. Med. Chem.* **2003**, *3*, 525–540. See also Chapter 8 in ref. 11c.

(11) (a) Stütz, A. E. *Iminosugars as Glycosidase Inhibitors: Nojirimycin and Beyond*; Wiley-VCH: Weinheim, 1999. (b) For a review of iminosugar-based glycosyl transferase inhibitors, see: Compain, P.; Martin, O. R. *Curr. Top. Med. Chem.* **2003**, *3*, 541–560. (c) Compain, P.; Martin, O. R. *Iminosugars: From Synthesis to Therapeutic Application*; Wiley: Chichester, 2007.

(12) Boucheron, C.; Desvergnès, V.; Compain, P.; Martin, O. R.; Lavi, A.; Mackeen, M.; Dwek, R. A.; Wormald, M.; Butters, T. D. *Tetrahedron Asymmetry* **2005**, *16*, 1747–1756.

(13) (a) Lee, R. E.; Smith, M. D.; Nash, R. J.; Griffiths, R. C.; McNeil, M.; Grewal, R. K.; Yan, W.; Besra, G. S.; Brennan, P. J.; Fleet, G. W. J. *Tetrahedron Lett.* **1997**, *38*, 6733–6736. (b) Pham-Huu, D.-P.; Gizaw, Y.; BeMiller, J. N.; Petrus, L. *Tetrahedron* **2003**, *59*, 9413–9417.

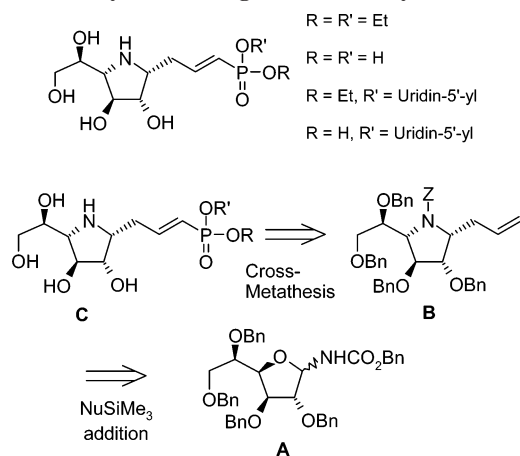
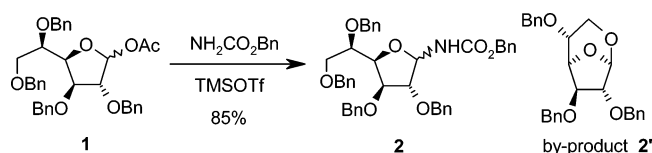
(14) Linkage by a diamide tether: Lee, R. E.; Smith, M. D.; Pickering, L.; Fleet, G. W. J. *Tetrahedron Lett.* **1999**, *40*, 8689–8692.

(15) (a) Liautard, V.; Christina, A. E.; Desvergnès, V.; Martin, O. R. *J. Org. Chem.* **2006**, *71*, 7337–7345. See also: (b) Desvergnès, S.; Desvergnès, V.; Martin, O. R.; Itoh, K.; Liu, H. W.; Py, S. *Bioorg. Med. Chem.* **2007**, *15*, 6443–6449.

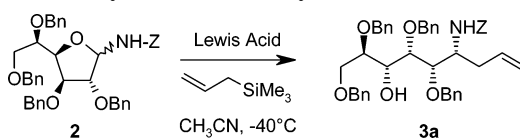
(16) For a preliminary communication, see: Liautard, V.; Desvergnès, V.; Martin, O. R. *Org. Lett.* **2006**, *8*, 1299–1302.

(17) (a) Sugiura, M.; Kobayashi, S. *Org. Lett.* **2001**, *3*, 477–480. (b) Sugiura, M.; Hagio, H.; Hirabayashi, R.; Kobayashi, S. *Synlett* **2001**, 1225–1228. (c) Sugiura, M.; Hagio, H.; Hirabayashi, R.; Kobayashi, S. *J. Am. Chem. Soc.* **2001**, *123*, 12510–12517. (d) Sugiura, M.; Hirabayashi, R.; Kobayashi, S. *Helv. Chem. Acta* **2002**, *85*, 3678–3691.

SCHEME 2. Synthetic Targets and Retrosynthesis

SCHEME 3. Synthesis of the *N*-Z Glucosylamine 2

SCHEME 4. Synthesis of the Allylamine 3a



Results and Discussion

1-*O*-Acetyl-2,3,5,6-tetra-*O*-benzyl-*D*-glucofuranose **1** was prepared from *D*-glucose on a 20 g scale by a four-step procedure.¹⁵ Under the conditions reported by Kobayashi (*Z*-NH₂, TMSOTf), the corresponding *N*-*Z* glucosylamine was obtained as a single product in 85% yield using 2 equiv of benzyl carbamate. When only 1.5 equiv or less of the carbamate was used, **2** was obtained as a mixture (from 96/4 to 84/16) with the byproduct **2'** (1,6-anhydro-2,3,5-tri-*O*-benzyl- β -*D*-glucofuranose)¹⁸ arising from a Lewis acid-assisted regioselective de-*O*-benzylation at O-6 (Scheme 3). It is noteworthy that the reaction also works directly from 2,3,5,6-tetra-*O*-benzyl-*D*-glucofuranose. This reaction was recently extended to other deactivated amines (e.g., TsNH₂) and other sugar hemiacetals.¹⁹

The glucosylamine **2** (α,β mixture) was then submitted to the Lewis acid-catalyzed reaction with allyltrimethylsilane (Scheme 4). After extensive investigations on the nature of the Lewis acid and on reaction conditions (Table 1), it was found that TMSOTf was the best reagent. 1.25 equiv of TMSOTf and 7 equiv of allyltrimethylsilane (added in portions) were necessary for the reaction to go to completion (entry 4). Compound **3a** was then isolated in a good yield of 86% and as a single *syn*-stereoisomer (see below).

Silylated Lewis acids and bismuth triflate, known to be somewhat azaphilic,²⁰ led to the best results. On the other hand,

TABLE 1. Allylation of **2** under Various Conditions^a

entry	lewis acid (equiv)	reaction time (h)	yield (%)
1	TMSOTf (0.1)	48	61
2	TMSOTf (0.5)	65	74
3	TMSOTf (1)	48	83
4	TMSOTf (1.25)	72	86
5	Bi(OTf) ₃ (0.1)	48	52
6	Bi(OTf) ₃ (0.5)	64	80
7	Bi(OTf) ₃ (1.25)	48	81
8	TIPSOTf (0.1)	48	72
9	TIPSOTf (1)	64	80
10	TIPSOTf (1.25)	48	80
11	HNTf ₂ (0.1)	48	36
12	HNTf ₂ (0.5)	48	75
13	HNTf ₂ (1.25)	48	53
14	Sc(OTf) ₃ (1.25)	48	0
15	BF ₃ ·Et ₂ O (1.25)	48	8

^a Seven equivalents of AllSiMe₃ were used in MeCN at -40 °C.

with the rather oxophilic BF₃ etherate, disappointing results were obtained, that could be linked to the numerous chelation sites (OBn, CO₂Bn) on the substrate, thereby decreasing the availability of the Lewis acid to form the *N*-acyl iminium ion (entry 15). Surprisingly, scandium triflate, which was expected to behave like bismuth triflate, did not lead to the conversion of the *N*-glucosylamine **2** whatever the reaction conditions used (entry 14). HNTf₂ is a very reactive acid but its air and moisture sensitivity limited the reproducibility of the reactions.

As a consequence, TMSOTf was used to scale up the synthesis of **3a** and to perform the addition of a variety of other silylated nucleophiles to *N*-glucofuranosylamine **2**, thus extending the synthetic value of this reaction (Scheme 5 and Table 2). Several useful functionalities could thus be introduced: with TMSCN, the reaction led to heptonitrile **3b** in an unoptimized 63% yield (entry 2). With silyl enol ethers, various keto functions were appended (entries 3, 4 and 5): a benzoylmethyl group with the silyl enol ether of acetophenone, a 2-oxocyclohexyl and a 2-oxocyclopentyl group with the silyl enol ethers of cyclohexanone and cyclopentanone respectively. It is noteworthy that a single stereoisomer was formed at the alkylation site α to the carbonyl group in cyclohexanone derivative **3d** whereas a mixture of two stereoisomers (ratio 2:1) was isolated for its cyclopentanone analog **3e**. A similar result was obtained using 2-(trimethylsiloxy)furan as the reagent: in this case, the two epimers at the alkylation site C₇, butenolides **3f** and **3f'** (27:73), could be isolated and fully characterized. These compounds are of particular interest as synthetic precursors of disaccharide mimicks (entry 6). Despite a disappointing yield (12%) and low stereoselectivity, it was possible to obtain the protected α -amino phosphonate **3g** by reaction with diethyl trimethylsilyl phosphite (entry 7). Unsaturated aliphatic chains could also be introduced as alkyne or allene moieties in moderate yields using 3-trimethylsilyl-1,2-butadiene or propargyltrimethylsilane as reagents (entries 8 and 9). The reaction of **2** with TMS isocyanate provided in one step the original cyclic carbamate **3j** resulting from the reaction of the free 4-hydroxyl group with the isocyanate function introduced as nucleophile. This stereochemically homogeneous structure incorporates an unprecedented, stable aminal of *D*-glucose (entry 10).

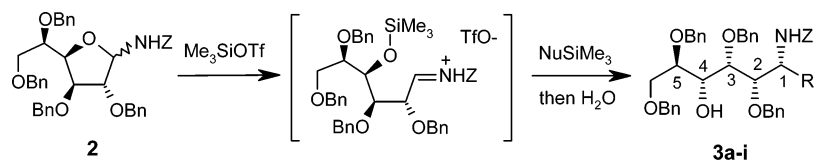
With one exception, all addition reactions exhibited a high degree of stereoselectivity: for the reactions leading to a single new stereogenic center by formation of a C–C bond, only one product was isolated (d.e. > 98%), which was shown, after cyclization, to have a 1,2-*syn* relationship, as expected from a

(18) Köll, P.; John, H.-G.; Schulz, J. *Liebigs Ann. Chem.* **1982**, 613–625.

(19) Liautard, V.; Pillard, C.; Desvergnès, V.; Martin, O. R. *Carbohydr. Res.* in press; doi 10.1016/j.carres.2007.11.022.

(20) Kobayashi, S.; Busujima, T.; Nagayama, S. *Chem.–Eur. J.* **2000**, 6, 3491–3494.

SCHEME 5. Addition of Silylated Nucleophiles to Glycosylamine 2

TABLE 2. Reactions of 2 with Silylated Nucleophiles NuSiMe₃^a

Entry	NuSiMe ₃	Reaction time	Product	Yield (%)
1		65h		74
2	TMSCN	56h		63
3		72h		81
4		68h		68
5		48h		72
6		44h		52
7	TMSOP(OEt) ₂	71h		12
8		60h		41
9		60h		54
10	TMSNCO	72h		58

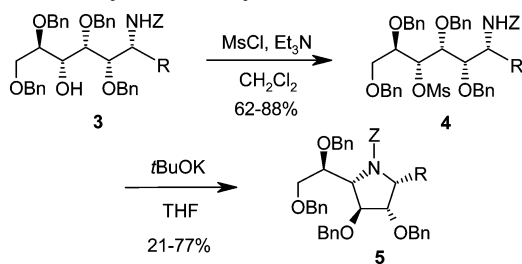
^a All reactions were performed using 0.5 equiv of TMSOTf and 7 equiv of silylated nucleophile, in CH₃CN at -40 °C.

avored Re-face addition on the intermediate iminium ion.¹⁴ The addition of the P-nucleophile, (EtO)₂POTMS, is the only reaction that proceeded with poor diastereoselectivity.

Next, to achieve cyclization, a classical two-step sequence involving mesylation of the free 4-OH group and subsequent treatment of the mesylate with a base (*t*-BuOK) provided the

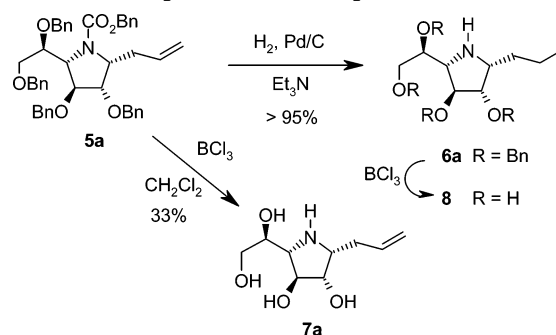
corresponding pyrrolidinol derivatives (series 5) in modest to good yields (Scheme 6 and Table 3). In particular, the nitrile 5b was found to be sensitive to base and we experienced some difficulties with the synthesis of this compound. Also, in most cases, better yields could be reached if the two steps from 3 were performed without isolating and purifying the mesylated

SCHEME 6. Synthesis of Pyrrolidinols 5



intermediate **4**. All new compounds **5a-j** are original imino-*C*-glycosyl mimicks of α -galactofuranosides. The functional groups in the aglycone can be used to tether a UDP surrogate by various synthetic approaches or to prepare analogs of other GalF conjugates.

Since the carbamate rotamers in compounds of series **5** complicated their analysis by NMR, the determination of

SCHEME 7. *N*-Deprotection of Compound 5a

configuration of the pseudo-anomeric center was performed on the *N*-deprotected pyrrolidine **6a** (Scheme 7) and **6b**.

Vicinal coupling constants as well as clear NOESY connectivities established unambiguously the 1,2-*cis* configuration of

TABLE 3. Synthesis of Pyrrolidinols 5

Entry	 4	Yield of 5 (%)	Product 5
1	4a R = CH ₂ CH=CH ₂	55 (70) ^a (74) ^b	 5a
2	4b R = CN	67 (69) ^b	 5b
3	4c R = CH ₂ COPh	21 (40) ^a	 5c
4	4d R =	36	 5d^c
5	4e R =	25	 5e^d
6	4h R =	63 (32) ^b	 5h
7	4i R =	77	 5i

^a Two-step sequence starting from **3**. ^b Three-step sequence starting from **2**. ^c Undetermined configuration at C*. ^d d.r. 67:33 at C*.

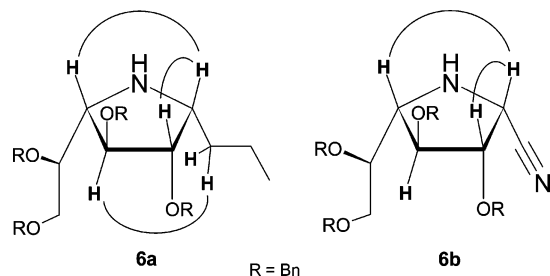


FIGURE 1. NOE correlations in compounds **6a** and **6b**.

the 1-*C*-substituted iminogalactitol derivative **6a** and **6b** and hence the 1,2-*syn* configuration of the addition product **3a** (Figure 1). By analogy and on the basis of similarities in NMR spectra of compounds in the same series, the 1,2-*cis* configuration was assigned to all pyrrolidinol derivatives **5** and the 1,2-*syn* configuration to all addition products **3**.

Further elaboration into UDP-Galf mimicks was performed from the α -*C*-allylated pyrrolidinol **5a** and functionalized alkenes by cross-metathesis. This powerful coupling process is relatively insensitive to the presence of heteroatoms in the reaction partners, with the exception of amino groups.²¹ It has already been used by our group²² and others²³ to access functionalized *C*-glycosyl compounds in iminosugars carrying a deactivating protecting group at nitrogen. The coupling procedure was first investigated using diethyl vinylphosphonate (DEVP). Hoveyda-Grubbs II complex [Ru]-**3**²⁴ turned out to be the most efficient catalyst and provided the cross-coupling product **9** in good yield, as the (*E*)-stereoisomer only, together with some homodimerization product **10** (10%). With Nolan's catalyst [Ru]-**2**,²⁵ the reaction did not go to completion and, surprisingly, no conversion was observed with Grubbs II complex [Ru]-**1** (Scheme 8 and Table 4).

Moreover, the latter compound (**10**) was obtained efficiently (53% yield) via the homo-cross-coupling of **5a** using 10 mol % of [Ru]-**3** as catalyst (Scheme 8). This compound is the precursor of interesting structures mimicking fragments of galactan. The coupling products **9** and **10** were *N*- and *O*-deprotected efficiently without affecting the alkene function using a large excess of BCl₃, thus providing unsaturated phosphonate **11** and the bis-imino-*C*-disaccharide **13** as hydrochlorides in 74% and 96% yield respectively. A further treatment of **11** using BrSiMe₃²⁶ led to the free phosphonic acid **12** (as hydrobromide) and catalytic hydrogenation of **13** provided compound **14**. These new imino-*C*-glycosyl compounds are of interest as potential inhibitors of various carbohydrate-processing enzymes.

SCHEME 8. Cross-Metathesis Reactions of **5a** and Deprotections

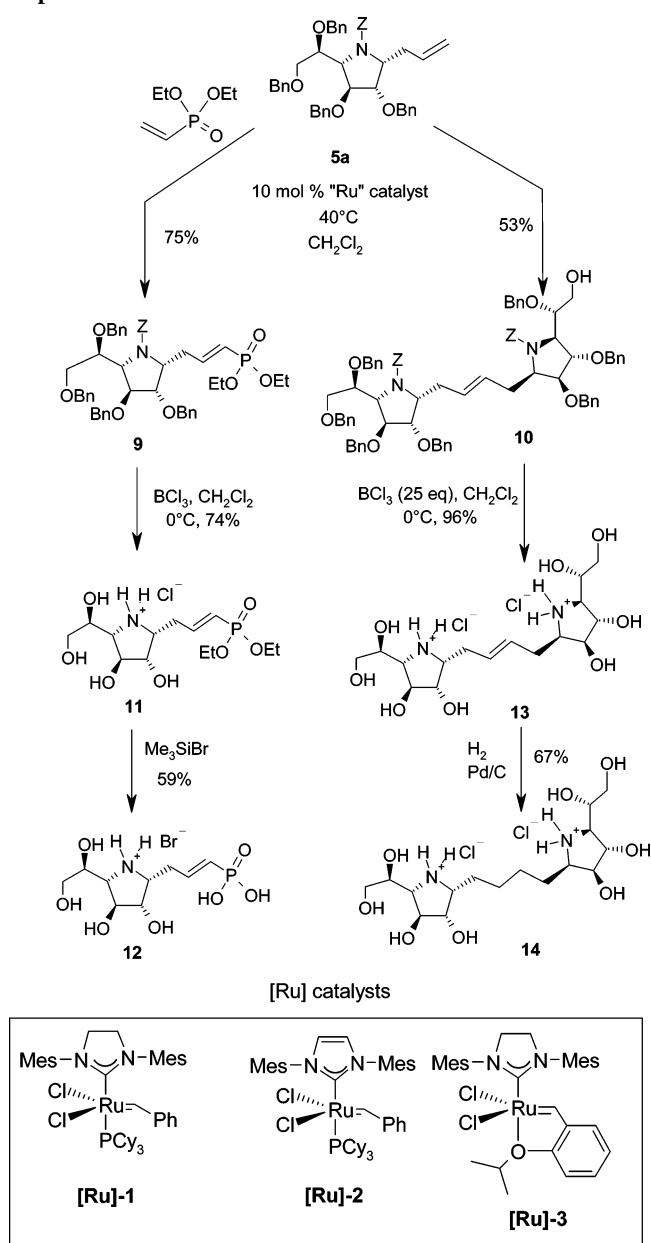


TABLE 4. Screening of Catalysts for Coupling of **5a** with DEVP

[Ru] catalyst	5a	9	10
Grubbs II [Ru]- 1	100%	—	—
Nolan [Ru]- 2	9%	68%	8%
Hoveyda-Grubbs II [Ru]- 3	—	75%	10%

Interestingly, the allyl group of **5a** could be isomerized into a 1-propenyl substituent to give compound **15** (69%) in the presence of Hoveyda's catalyst when the reaction was performed in methanol at higher temperature, as shown recently by Hanessian and co-workers.²⁷ The same isomerization process was also successful (in 61% yield) using Pd(MeCN)₂Cl₂ (10 mol % in toluene at 60 °C for 48 h).²⁸ Cross-metathesis with

(27) Hanessian, S.; Giroux, S.; Larsson, A. *Org. Lett.* **2006**, *24*, 5482–5484.

(28) Chang, G. X.; Lowary, T. *Tetrahedron Lett.* **2006**, *47*, 4561–4564.

(21) Compain, P. *Adv. Synth. Catal.* **2007**, *349*, 1829–1846.

(22) Godin, G.; Compain, P.; Martin, O. R. *Org. Lett.* **2003**, *5*, 3269–3272.

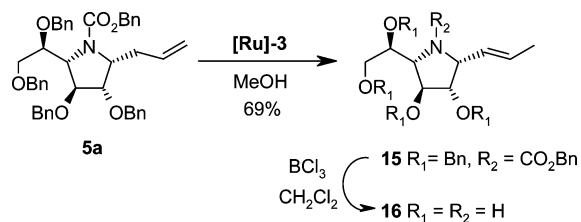
(23) Dondoni, A.; Giovannini, P. P.; Perrone, D. *J. Org. Chem.* **2005**, *70*, 5508–5518.

(24) Kingsbury, J. S.; Harrity, J. P. A.; Bonitatebus, P. J.; Hoveyda, A. H. *J. Am. Chem. Soc.* **1999**, *121*, 791–799. See also: Vehlou, K.; Maechling, S.; Blechert, S. *Organometallics* **2006**, *25*, 25–28.

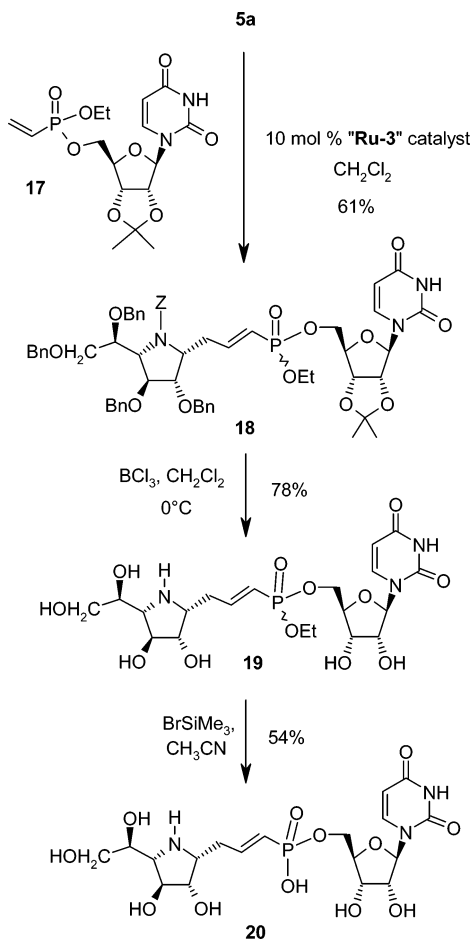
(25) (a) Huang, J.; Stevens, E. D.; Nolan, S. P.; Petersen, J. L. *J. Am. Chem. Soc.* **1999**, *121*, 2674–2678. (b) Amblard, F.; Nolan, S. P.; Schinazi, R. F.; Agrofoglio, L. A. *Tetrahedron* **2005**, *61*, 537–544. For an application to a homodimerization process, see, e.g., Grellepois, F.; Crousse, B.; Bonnet-Delpon, D.; Bégue, J. P. *Org. Lett.* **2005**, *7*, 5219–5222.

(26) For phosphonate deprotection using BrSiMe₃, see, e.g.: (a) Vidil, C.; Morère, A.; Garcia, M.; Barragan, V.; Hamdaoui, B.; Rochefort, H.; Montero, J.-L. *Eur. J. Org. Chem.* **1999**, 447–450. (b) Ladame, S.; Fauré, R.; Denier, C.; Lakhdar-Ghazal, F.; Wilson, M. *Org. Biomol. Chem.* **2005**, *3*, 2070–2072.

SCHEME 9. Isomerization of Allyl Group in 5a



SCHEME 10. Synthesis of UDP-GalF Mimick 20



other alkenes however was unsuccessful from **15**. Deprotection of **15** by the action of excess BCl_3 provided **16** (Scheme 9).

The same coupling approach was used to obtain the α -linked UDP-GalF mimicks **20** using the ethyl uridin-5'-yl vinylphosphonate **17**¹⁵ as cross-metathesis partner (Scheme 10). As with the previous reagent, total conversion of compound **5a** was achieved using Hoveyda-Grubbs II complex **[Ru]-3** as the catalyst. The iminosugar-nucleotide conjugate **18** was thus obtained as a mixture of non-separable P^* -epimers in 61% yield together with a small amount of compound **10** (13%). A two-step deprotection sequence led to the fully deprotected target compound **20** (42% overall). Treatment of **18** with BCl_3 in excess at 0°C promoted the complete deprotection of the ribofuranose and iminogalactitol moieties and led to the phosphonic diester **19**. The ethyl phosphonate could then be cleaved selectively by reacting **19** with BrSiMe_3 to provide UDP-GalF analog **20**. This compound incorporates an iminosugar moiety that mimicks the putative galactofuranosyl cation

TABLE 5. Inhibition of *E. Coli* UGM by Selected Compounds^a

compound #	inhibitor concentration	residual activity
7a	25 mM	61%
8	2.5 mM	52.1%
13	2.5 mM	50%
14	2.5 mM	36.9%
16	2.5 mM	59.2%
19	25 mM	52%
20	2.5 mM	43%

^a See experimental procedures in ref 15b.

involved in the reactions catalyzed by UDP-Gal mutase and/or UDP-GalF transferases and the three-carbon linker places the nucleotide fragment at an appropriate distance with respect to the pseudosugar moiety. To the best of our knowledge this compound is the closest iminosugar-based analog of UDP-GalF ever reported.

The inhibitory activity of compounds **7a**, **8**, **13**, **14**, **16**, **19**, and **20** toward UGM from *Escherichia coli* was then evaluated and the results are reported in Table 5. Inhibition assays on the purified enzyme were conducted in the reverse direction in which UDP-GalF is converted to UDP-Galp using an HPLC method developed by Lee and co-workers²⁹ under the same conditions as previously reported.^{15b} Residual activity is expressed in % of the activity in absence of inhibitor.

All derivatives tested were found to exhibit weak inhibition of UGM. Residual activity is in the order of 50% for most compounds at 2.5 mM inhibitor concentration. Noticably, the presence of the UMP motif, which was anticipated to provide significant binding, increased only to a very minor extent the activity of the compounds carrying a simple 3-carbon substituent at C-1. This is even more surprising in view of Kiessling's recent work on chemical probes for UGM which suggested that the presence of an aromatic group mimicking uracil is essential for better binding with the enzyme.³⁰ Most of the recognition by the enzyme appears to arise from the 1,4-iminogalactitol moiety, and dimerization of this unit, as in **13** and **14**, contributes to enhance the inhibitory activity. Compound **14** is indeed the most active inhibitor of the series.

In conclusion, we report in this article a convenient and highly stereoselective methodology for the synthesis of a diversity of α -1-C-substituted-1,4-dideoxy-1,4-imino-D-galactitols; such compounds constitute novel, original galactofuranoside mimicks. One of these compounds was converted into UDP-GalF analogs (**19**, **20**) in which UMP is linked to the galacto-pyrrolidinol by a 3-C tether, thereby mimicking closely the structure of the natural sugar nucleotide; carbon-linked disaccharide analogs (**13**, **14**) were also generated from the same precursor. Whereas the activity of compounds such as **19** and **20** as inhibitors of UGM was found to be weak, these sugar nucleotide analogs remain promising chemical probes of the GalF transfer process involved in the biosynthesis of mycobacterial galactans. In addition the 1-C-substituted iminogalactitol derivatives and the corresponding dimers are of interest as potential inhibitors of various glycofuranosidases. Further biological investigations on these compounds are in progress and the results will be reported in due course.

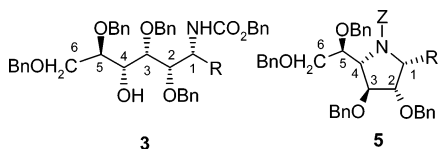
(29) Lee, R.; Monsey, D.; Weston, A.; Duncan, K.; Rithner, C.; McNeil, M. *Anal. Biochem.* **1996**, *242*, 1–7.

(30) Carlsson, E. E.; May, J. F.; Kiessling, L. L. *Chem. Biol.* **2006**, *13*, 825–837.

Experimental Section

General Procedure A: Addition of Silylated Nucleophiles.

In a dry 10 mL flask, under Ar, the silylated nucleophile (NuSiMe₃ in Table 2) (1.45 mmol, 7 equiv) was added to a solution of *N*-benzyloxycarbonyl-2,3,5,6-tetra-*O*-benzyl- α,β -D-glucopyranosylamine **2** (140 mg, 0.21 mmol)¹⁹ in anhydrous acetonitrile (2.1 mL). The mixture was stirred at -40 °C during 15 min and trimethylsilyl triflate (19 μ L, 0.10 mmol, 0.5 equiv) was then added by portions to the colorless solution. The mixture was stirred at -40 °C during 20–82 h, and the reaction was then quenched with saturated aqueous NaHCO₃ (1 mL). Ethyl acetate (20 mL) was added and the organic layer was separated, washed with brine (20 mL) and dried over sodium sulfate. The solvents were removed by evaporation under vacuum to afford a brown oil which was purified by flash chromatography (toluene/acetone 99:1 to 98:2 containing a trace of Et₃N unless otherwise indicated).



1(R)-1-C-Allyl-2,3,5,6-tetra-*O*-benzyl-1-benzyloxycarbonylamino-1-deoxy-D-glucitol (3a). According to general procedure A, compound **3a** was obtained as a colorless oil (129 mg, 86%) from **2** (140 mg, 0.21 mmol) after a stirring time of 72 h. [α]_D²⁵ -6 (*c* = 1.22, CHCl₃); IR (NaCl, cm⁻¹) 3436, 3029, 3013, 2927, 2866, 1716, 1498, 1454, 1333, 1216, 1097, 1060, 1027; ¹H NMR (500 MHz, CDCl₃) δ 2.18–2.31 (m, 2H, CH₂CH=CH₂), 2.56 (d, 1H, *J* \approx 9 Hz, OH), 3.55–3.59 (m, 1H, H4), 3.62–3.68 (m, 2H, H5, H6a), 3.72 (d, 1H, *J* \approx 8 Hz, H2), 3.81–3.91 (m, 3H, H1, H3, H6b), 4.22 (d, 1H, *J* \approx 11 Hz, OCH₂Ph), 4.29 (d, 1H, *J* \approx 11.5 Hz, OCH₂Ph), 4.43–4.49 (m, 3H, OCH₂Ph), 4.63 (d, 2H, *J* \approx 11.5 Hz, OCH₂Ph), 4.73 (d, 1H, *J* \approx 11 Hz, OCH₂Ph), 4.86–4.96 (m, 3H, OCH₂Ph, CH₂=CH), 5.04 (d, 1H, *J* \approx 12.5 Hz, OCH₂Ph) 5.13 (d, 1H, *J* \approx 9.5 Hz, NH), 5.62–5.70 (m, 1H, CH=CH₂), 7.08–7.26 (m, 25H, CH_{Ar}s); ¹³C NMR (125 MHz, CDCl₃) δ 38.3 (CH₂-CH=CH₂), 51.5 (C1), 66.8 (OCH₂Ph), 70.9 (C5), 71.2 (C6), 71.7, 73.7, 74.7, and 75.2 (OCH₂Ph), 78.2 (C3+C4), 79.7 (C2), 117.9 (CH₂=CH), 127.5–128.6 (CH_{Ar}s), 134.7 (CH=CH₂), 136.7, 138.3, 138.4, and 138.6 (C_{qAr}), 156.1 (C=O); (+) MS (ESI): *m/z* = 733.5 [M + NH₄]⁺, 738.5 [M + Na]⁺; HRMS (ESI): calcd for C₄₅H₄₉NNaO₇: *m/z* = 738.3407 [M + Na]⁺, found: *m/z* = 738.3401. Anal. Calcd. for C₄₅H₄₉NO₇: C, 75.50; H, 6.90; N, 1.96. Found: C, 75.72; H, 6.96; N, 1.84.

3,4,6,7-Tetra-*O*-benzyl-2-benzyloxycarbonylamino-2-deoxy-D-glycero-D-ido-heptono-nitrile (3b). According to general procedure A, compound **3b** was obtained as a colorless oil (35.1 mg, 63%) from **2** (53.4 mg, 0.079 mmol) using 0.5 equiv of trimethylsilyl triflate and after a stirring time of 56 h. [α]_D²⁵ -4 (*c* = 0.45, CHCl₃); IR (NaCl, cm⁻¹) 3427, 3033, 2253, 1726, 1497, 1455, 1271, 1215, 1096, 1028; ¹H NMR (500 MHz, CDCl₃) δ 2.70 (broad s, 1H, OH), 3.57–3.61 (m, 1H, H4), 3.65–3.73 (m, 1H, H6a), 3.78–3.82 (m, 2H, H6b + H5), 3.85 (d, 1H, *J* \approx 7 Hz, H3), 4.03–4.05 (m, 1H, H2), 4.28 (t, 2H, *J* \approx 10.5 Hz, OCH₂Ph), 4.50–4.65 (m, 4H, OCH₂Ph), 4.71 (AB, 2H, *J* \approx 11 Hz, OCH₂Ph), 4.91–4.93 (m, 1H, H-1), 5.02–5.12 (m, 2H, OCH₂Ph), 5.67–5.69 (m, 1H, NH), 7.14–7.32 (m, 25H, CH_{Ar}s); ¹³C NMR (125 MHz, CDCl₃) δ 43.8 (C1), 67.9 (OCH₂Ph), 70.2 (C6, C5), 71.8, 73.7, 74.5, and 75.0 (OCH₂Ph), 76.6 (C3), 77.6 (C4), 78.0 (C2), 118.2 (CN), 127.8–130.0 (CH_{Ar}s), 135.8, 136.8, 137.7, 138.0, and 138.3 (C_{qAr}), 155.4 (C=O); (+) MS (ESI): *m/z* = 719 [M + NH₄]⁺; HRMS (ESI) calcd for C₄₃H₄₄N₂NaO₇: *m/z* = 723.3046 [M + Na]⁺, found: *m/z* = 723.3041.

4,5,7,8-Tetra-*O*-benzyl-3-benzyloxycarbonylamino-2,3-dideoxy-1-C-phenyl-D-glycero-D-ido-1-octulose (3c). According to general procedure A, compound **3c** was obtained as a colorless oil (200

mg, 81%) from **2** (208 mg, 0.31 mmol), using 0.5 equiv of trimethylsilyl triflate and after a stirring time of 72 h. [α]_D²⁵ -3 (*c* = 0.69, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 2.93 (d, 1H, *J* \approx 7.5 Hz, OH), 3.01–3.05 (m, 1H, CH₂COPh), 3.19–3.26 (m, 1H, CH₂COPh), 3.73–3.79 (m, 2H, H2 + H6a), 3.92–3.95 (m, 4H, H6b + H5 + H3 + H4), 4.33–4.49 (m, 5H, H1 + OCH₂Ph), 4.55 (broad s, 2H, OCH₂Ph), 4.70–4.77 (m, 2H, OCH₂Ph), 5.02 (d, 1H, *J* \approx 12 Hz, OCH₂Ph), 5.07 (d, 1H, *J* \approx 12 Hz, OCH₂Ph), 5.43–5.47 (m, 1H, NH), 7.08–7.81 (m, 30H, H_{Ar}s); ¹³C NMR (125 MHz, CDCl₃) δ 41.5 (CH₂COPh), 48.3 (C1), 66.9 (OCH₂Ph), 70.3 (C5), 71.2 (C6), 71.7, 73.6, 74.4, and 75.0 (OCH₂Ph), 77.9 (C3 or C4), 78.3 (C2), 78.7 (C4 or C3), 127.5–128.6, 133.3, 136.5, 138.0, 138.3, 138.4, and 138.8 (C_{qAr}), 155.9 (C=O), 198.4 (C=O); HRMS (ESI) calcd for C₅₀H₅₁NNaO₈: *m/z* = 816.3512 [M + Na]⁺; found: *m/z* = 816.3506.

1(R)-2,3,5,6-Tetra-*O*-benzyl-1-benzyloxycarbonylamino-1-deoxy-1-C-(2-oxocyclohexyl)-D-glucitol (3d). According to general procedure A, compound **3d** (107 mg, 68%) was obtained from compound **2** (138.1 mg, 0.205 mmol) as a single product and as a colorless oil after purification by flash chromatography (EP/AcOEt 4:1). [α]_D²⁵ -19 (*c* = 0.99, CHCl₃); IR (NaCl, cm⁻¹) 3435, 3063, 3031, 2935, 2863, 1718, 1702, 1498, 1454, 1210, 1072, 1028; ¹H NMR (250 MHz, CDCl₃) δ 1.48–2.10 (m, 7H), 2.17–2.33 (m, 1H), 2.78–3.00 (m, 2H, OH and CHCO), 3.57–3.75 (m, 2H, H5 + H6a), 3.77–3.90 (m, 2H, H3 + H6b), 3.92–4.06 (m, 2H, H2 + H4), 4.08–4.18 (m, 1H, H1), 4.24 (d, 1H, *J* \approx 11 Hz, OCH₂Ph), 4.31 (d, 1H, *J* \approx 11.5 Hz, OCH₂Ph), 4.43–4.59 (m, 4H, OCH₂Ph), 4.60–4.72 (m, 2H, OCH₂Ph), 5.52 (d, 1H, *J* \approx 10 Hz, NH), 7.06–7.40 (m, 25H, CH_{Ar}s); ¹³C NMR (125 MHz, CDCl₃) δ 24.4, 27.8, 31.3, 42.4, and 52.1 (cyclohexyl CH₂ and CH), 53.5 (C1), 66.8 (OCH₂Ph), 70.0 (C4), 71.0 (C6), 71.9, 73.6, 73.7, and 74.0 (OCH₂Ph), 77.0 (C3), 77.3 (C2), 78.6 (C5), 127.5–128.8 (CH_{Ar}s), 136.8, 138.0, 138.2, 138.3, and 138.7 (C_{qAr}), 156.8 (C=O), 211.7 (C=O); (+) MS (ESI): *m/z* = 789.5 [M + NH₄]⁺, 795.0 [M + Na]⁺; HRMS (ESI) calcd for C₄₈H₅₃NNaO₈: *m/z* = 794.3669 [M + Na]⁺; found: *m/z* = 794.3672.

1(R)-2,3,5,6-Tetra-*O*-benzyl-1-benzyloxycarbonylamino-1-deoxy-1-C-(2-oxocyclopentyl)-D-glucitol (3e). According to general procedure A, compound **3e** (367.5 mg, 70%) was obtained from compound **2** (463 mg, 0.688 mmol) as a colorless oil (mixture of diastereoisomers at *cp*-C2) after purification by flash chromatography (EP/ AcOEt 5:2 to 2:1). IR (NaCl, cm⁻¹) 3435, 3063, 3017, 1720, 1454, 1404, 1217, 1093, 1028; ¹H NMR (400 MHz, CDCl₃) δ 1.22–2.42 (m, 7H), 2.71 (br s, 1H, OH), 3.59–4.05 (m, 6H), 4.20–4.95 (m, 8H, OCH₂Ph and H2), 5.00–5.42 (m, 2H, OCH₂Ph and NH), 7.11–7.42 (m, 25H, CH_{Ar}s); ¹³C NMR (100 MHz, CDCl₃; d1 and d2 for diast. 1 and 2, respectively; *cp* for cyclopentyl) δ 20.3 (d1, *cp*-C4), 20.5 (d2, *cp*-C4), 27.3 (d1, *cp*-C3), 27.7 (d2, *cp*-C3), 38.7 (d1, *cp*-C5), 38.9 (d2, *cp*-C5), 50.2 (d2, *cp*-C2), 50.9 (d1, *cp*-C2), 51.2 (d1, C1), 52.3 (d2, C1), 67.0 (OCH₂Ph), 70.3 (d1, C4), 70.7 (d2, C4), 71.0 (d2, C6), 71.5 (d1, C6), 71.7, 71.8, 73.58, 73.63, 74.2, 74.38, 74.44, and 74.8 (OCH₂Ph), 77.4 (C3), 77.5 (C3), 77.9 (d1, C5), 78.1 (d2, C5), 78.3 (d1, C2), 78.5 (d2, C2), 127.4–128.5 (CH_{Ar}s), 136.6, 136.7, 137.9, 138.2, 138.3, 138.38, 138.40, 138.5, 138.6, and 138.9 (C_{qAr}), 156.3 (C=O), 156.7 (C=O), 218.9 (*cp*-C1), 219.2 (*cp*-C1); (+) MS (ESI): *m/z* = 775.5 [M + NH₄]⁺, 780.0 [M + Na]⁺; HRMS (ESI) calcd for C₄₇H₅₁NNaO₈: *m/z* = 780.3512 [M + Na]⁺; found: *m/z* = 780.3513.

1(R)-2,3,5,6-Tetra-*O*-benzyl-1-benzyloxycarbonylamino-1-C-but-2-ynyl-1-deoxy-D-glucitol (3h). According to general procedure A, compound **3h** (100 mg, 47%) was obtained from compound **2** (196 mg, 0.291 mmol) as a colorless oil after purification by flash chromatography (Toluene/ AcOEt 16:1). [α]_D²⁵ -6 (*c* = 0.71, CHCl₃); IR (NaCl, cm⁻¹) 3429, 3063, 3032, 2919, 2866, 1718, 1498, 1454, 1334, 1266, 1216, 1095, 1062, 1027; ¹H NMR (400 MHz, CDCl₃) δ 1.71 (s, 3H, CH₃), 2.30–2.57 (m, 2H, CH₂), 2.66 (br s, 1H, OH), 3.56–3.71 (m, 1H, H5), 3.73–3.82 (m, 1H, H4), 3.85–3.94 (m, 3H, 2H6 and H3), 3.98–4.07 (m, 1H, H1), 4.09–4.18 (m, 1H, H2), 4.23–4.42 (m, 2H, OCH₂Ph), 4.49–5.00 (m,

7H, OCH₂Ph), 5.07–5.17 (m, 1H, OCH₂Ph), 5.24 (d, 1H, $J \approx 9.6$ Hz, NH), 7.18–7.43 (m, 25H, CH_{Ar}'s); ¹³C NMR (100 MHz, CDCl₃) δ 3.6 (CH₃), 23.6 (CH₂), 51.2 (C1), 66.9 (OCH₂Ph), 70.5 (C4), 71.2 (C6), 71.7, 73.6, and 74.5 (OCH₂Ph), 75.36 (C_{alkyne}-CH₃), 75.41 (OCH₂Ph), 76.8 (C_{alkyne}-CH₂), 77.9 (C3), 78.2 (C5), 78.9 (C2), 127.5–128.6 (CH_{Ar}'s), 136.6, 138.2, 138.3, and 138.7 (Cq_{Ar}), 155.9 (C=O); (+) MS (ESI): $m/z = 728$ [M + H]⁺, 745.5 [M + NH₄]⁺, 750.5 [M + Na]⁺; HRMS (ESI) calcd for C₄₆H₄₉NNaO₇: $m/z = 750.3407$ [M + Na]⁺, found: $m/z = 750.3416$.

1(R)-1-C-Allenyl-2,3,5,6-tetra-O-benzyl-1-benzyloxycarbonylamino-1-deoxy-D-glucitol (3i). According to general procedure A, compound **3i** (149 mg, 54%) was obtained from compound **2** (259 mg, 0.233 mmol) as a colorless oil after purification by flash chromatography (PE/ AcOEt 7:1 to 6:1). [α]_D²⁵ –1.6 ($c = 3.14$, CHCl₃); IR (NaCl, cm⁻¹) 3432, 3063, 3031, 2926, 2867, 1958, 1719, 1498, 1455, 1399, 1337, 1276, 1215, 1096, 1062, 1028, 854; ¹H NMR (400 MHz, CDCl₃) δ 2.67 (d, 1H, $J \approx 8.8$ Hz, OH), 3.57–3.97 (m, 6H), 4.31 (d, 1H, $J \approx 11.2$ Hz, OCH₂Ph), 4.35 (d, 1H, $J \approx 11.8$ Hz, OCH₂Ph), 4.47–4.65 (m, 4H, OCH₂Ph and H1), 4.67–4.84 (m, 5H, OCH₂Ph and CH₂=), 4.98 (d, 1H, $J \approx 12$ Hz, OCH₂-Ph), 5.14 (d, 1H, $J \approx 12$ Hz, OCH₂Ph), 5.24 (q, 1H, $J \approx 5.9$ Hz, CH=), 5.38 (d, 1H, $J \approx 9.2$ Hz, NH), 7.17–7.38 (m, 25H, CH_{Ar}'s); ¹³C NMR (100 MHz, CDCl₃) δ 50.4 (C1), 66.9 (OCH₂Ph), 70.9 (C6), 71.7, 73.6, 74.8, and 75.4 (OCH₂Ph), 78.0 (CH), 78.6 (CH₂=), 81.5 (CH), 91.8 (CH=), 127.5–128.6 (CH_{Ar}'s), 136.6, 138.19, 138.21, 138.4, and 138.6 (Cq_{Ar}), 156.1 (C=O), 207.6 (C=C); (+) MS (ESI): $m/z = 714.5$ [M + H]⁺, 731.5 [M + NH₄]⁺, 736.5 [M + Na]⁺; HRMS (ESI) calcd for C₄₅H₄₇NNaO₇: $m/z = 736.3250$ [M + Na]⁺; found: $m/z = 736.3260$.

2,3,5,6-Tetra-O-benzyl-D-glucose aminal N_S-benzyl carbamate N_R-O-cyclic carbamate (3j). According to general procedure A, compound **3j** (96.5 mg, 58%) was obtained from compound **2** (157 mg, 0.233 mmol) as a colorless oil after purification by flash chromatography (PE/AcOEt 3:1). [α]_D²⁵ –26 ($c = 1.05$, CHCl₃); IR (NaCl, cm⁻¹) 3392, 3321, 3063, 3032, 2869, 1732, 1513, 1499, 1455, 1266, 1309, 1227, 1102, 1028, 1046; ¹H NMR (400 MHz, CDCl₃) δ 3.64 (t, 1H, $J \approx 3.6$ Hz, H2), 3.80 (dd, 1H, $J \approx 4.4$ and 10.4 Hz, H6a), 3.88 (ddd, 1H, $J \approx 1.6$, 4.4 and 9.6 Hz, H5), 3.99 (dd, 1H, $J \approx 1.6$ and 10.4 Hz, H6b), 4.09 (d, 1H, $J \approx 3.6$ Hz, H3), 4.25–4.39 (m, 3H, OCH₂Ph), 4.45–4.65 (m, 5H, OCH₂Ph and H4), 4.79 (d, 1H, $J \approx 11.6$ Hz, OCH₂Ph), 4.95–5.01 (m, 2H, OCH₂Ph and H1), 5.12 (d, 1H, $J \approx 12.4$ Hz, OCH₂Ph), 5.86 (d, 1H, $J \approx 6$ Hz, NH), 6.99 (d, 1H, $J \approx 9.2$ Hz, NH), 7.04–7.36 (m, 25H, CH_{Ar}'s); ¹³C NMR (100 MHz, CDCl₃) δ 61.0 (C1), 67.2 (OCH₂-Ph), 69.1 (C6), 71.0 (C2), 71.8, 72.0, 73.3, and 73.6 (OCH₂Ph), 74.9 (C4), 75.6 (C5), 76.0 (C3), 127.6–128.7 (CH_{Ar}'s), 136.1, 136.7, 137.0, 138.3, and 138.4 (Cq_{Ar}), 155.5 (C=O), 157.3 (C=O); (+) MS (ESI): $m/z = 717.5$ [M + H]⁺, 734.5 [M + NH₄]⁺, 739.5 [M + Na]⁺; HRMS (ESI) calcd for C₄₃H₄₄N₂NaO₈: $m/z = 739.2995$ [M + Na]⁺; found: $m/z = 739.2980$.

General Procedure B: Cyclization Procedure. In a dry flask (10 mL), under Ar, methanesulfonyl chloride (8.2 μ L, 0.11 mmol, 2.1 equiv) was added by portions to a solution of substrate **3** (38 mg, 0.053 mmol) in anhydrous dichloromethane (530 μ L) containing triethylamine (16 μ L, 0.12 mmol, 2.2 equiv). The yellow mixture was stirred at room temperature for 5–12 h. After total conversion (TLC monitoring), the reaction was quenched with saturated aqueous NH₄Cl (1 mL). Ethyl acetate (20 mL) was added, the organic layer was separated, washed with brine (20 mL) and dried over sodium sulfate. The solvents were then evaporated, thus providing the corresponding methanesulfonate **4** as a yellow oil which could be used in the next step without further purification.

Crude **4** (142 mg, 0.18 mmol) was dissolved in anhydrous THF (1.8 mL) and *t*-BuOK (20.12 mg, 0.18 mmol) was added. The mixture was stirred during 12–24 h at room temperature. After total conversion (TLC monitoring), the reaction was quenched with saturated aqueous NH₄Cl (1 mL). Ethyl acetate (20 mL) was added, the organic layer was separated, washed with brine (20 mL) and dried over sodium sulfate. Then the solvents were evaporated, thus

providing a colorless oil which was purified by flash chromatography (PE/AcOEt 7:1 containing a trace of Et₃N).

1(R)-1-C-Allyl-2,3,5,6-tetra-O-benzyl-N-benzyloxycarbonyl-1,4-dideoxy-1,4-imino-D-galactitol (5a). According to general procedure B applied to **3a**, compound **5a** was obtained as a colorless oil (70% yield, 2 steps), using 2.5 equiv ^tBuOK. [α]_D²⁵ + 7 ($c = 0.73$, CHCl₃); IR (NaCl, cm⁻¹) 3359, 3064, 3032, 2929, 2867, 1699, 1497, 1454, 1402, 1352, 1097, 1070, 1028; ¹H NMR (500 MHz, acetone-*d*₆) δ 2.38–2.45 (m, 1H, CH₂CH=CH₂), 2.65 (broad s, 1H, CH₂CH=CH₂), 3.62–3.64 (m, 1H, H6a), 3.74–3.77 (m, 1H, H6b), 3.97–4.01 (m, 1H, H5), 4.12 (dd, 1H, $J \approx 3$ Hz, $J \approx 6.5$ Hz, H2), 4.21–4.24 (m, 1H, H1), 4.26 (t, 1H, $J \approx 3$ Hz, H3), 4.30–4.32 (m, 1H, H4), 4.46 (d, 1H, $J \approx 12$ Hz, OCH₂Ph), 4.51 (d, 1H, $J \approx 12$ Hz, OCH₂Ph), 4.53–4.74 (m, 6H, OCH₂Ph), 4.94–4.96 (m, 1H, CH₂=CH), 5.01–5.05 (m, 1H, OCH₂Ph), 5.10 (d, 1H, $J \approx 12$ Hz, OCH₂Ph), 5.17 (d, 1H, $J \approx 12$ Hz, OCH₂Ph), 5.75–5.86 (m, 1H, CH=CH₂), 7.27–7.42 (m, 25H, CH_{Ar}'s); ¹³C NMR (125 MHz, acetone-*d*₆) δ 35.2 (CH₂CH=CH₂), 62.2–62.9 (C1), 66.2 (C4), 68.1 (OCH₂Ph), 72.7 (C6), 73.3, 73.7, 74.3, and 74.5 (OCH₂-Ph), 79.6 (C5), 82.6 (C3), 83.9 (C2), 117.4 (CH=CH₂), 128.8–131.0 (CH_{Ar}'s), 134.6 (Cq_{Ar}), 135.1 (Cq_{Ar}), 137.7 (CH=CH₂), 139.9, 139.8, 140.2, 140.4 and 140.7 (Cq_{Ar}), 158.0 (C=O); (+) MS (ESI): $m/z = 715.5$ [M + NH₄]⁺; HRMS (ESI) calcd for C₄₅H₄₇NNaO₆: $m/z = 720.3301$ [M + Na]⁺; found: $m/z = 720.3296$. Anal. Calcd. for C₄₅H₄₇NO₆: C, 77.45; H, 6.79; N, 2.01. Found: C, 77.80; H, 6.68; N, 2.09. Note: A sample of mesylate **4a** was purified (PE/AcOEt 85/15 containing a trace of Et₃N); yield: 62%. Then using the general cyclization procedure, compound **5a** was obtained in 55% yield.

3,4,6,7-Tetra-O-benzyl-N-benzyloxycarbonylamino-2,5-dideoxy-2,5-imino-D-glycero-L-gluco-heptononitrile (5b). According to general procedure A (using TMSCN) and B applied to **3b**, compound **5b** was obtained as a colorless oil in 69% yield (3-step yield). [α]_D²⁵ +17 ($c = 0.66$, CHCl₃); IR (NaCl, cm⁻¹) 3055, 2987, 2306, 1715, 1454, 1421, 1265, 1099, 1028; ¹H NMR (500 MHz, acetone-*d*₆) δ 3.65–3.67 (m, 1H, H6a), 3.71–3.73 (m, 1H, H6b), 4.05 (br s, 1H, H5), 4.30–4.31 (m, 2H, H3 + H4), 4.43–4.50 (m, 3, H2 + OCH₂Ph), 4.52 (s, 2H, OCH₂Ph), 4.63–4.78 (m, 3H, OCH₂Ph), 4.77 (d, 1H, $J \approx 11.8$ Hz, OCH₂Ph), 5.11 (very br s, 2H, OCH₂Ph), 5.30 (br s, 1H, H1), 7.21–7.42 (m, 25H, CH_{Ar}'s); ¹³C NMR (125 MHz, acetone-*d*₆) δ 53.9–54.2 (C1), 66.0 (C4), 69.0 (OCH₂Ph), 72.3–72.6 (C6), 73.0 (OCH₂Ph), 74.3 (OCH₂Ph), 74.6 (OCH₂Ph), 78.5 (C5), 82.7–82.9 (C2), 83.7–83.8 (C3), 117.8 (CN), 128.9–130.3 (CH_{Ar}'s), 138.2, 139.1, 139.7, 140.3, and 140.5 (Cq_{Ar}); (+) MS (ESI): $m/z = 700.5$ [M + NH₄]⁺; HRMS (ESI) calcd for C₄₃H₄₂N₂NaO₆: $m/z = 705.2941$ [M + Na]⁺; found: $m/z = 705.2944$.

Note: Stepwise process from compound **3b** (75.9 mg, 0.108 mmol): mesylated intermediate **4b** was obtained as a colorless oil (74.5 mg, 88%) after purification by flash chromatography (PE/AcOEt 3:1). Compound **5b** was then obtained from **4b** (173 mg, 0.222 mmol) as a colorless oil (100.7 mg, 67%) after purification by flash chromatography (PE/AcOEt 85:15).

4,5,7,8-Tetra-O-benzyl-N-benzyloxycarbonylamino-2,3,6-trideoxy-3,6-imino-1-C-phenyl-D-glycero-L-gluco-1-octulose (5c). Compound **5c** was prepared from **3c** according to general procedure B; mesylation was immediately followed by the cyclization step because intermediate **4c** degrades rapidly. Compound **5c** was obtained as a colorless oil (40%, 2 steps). [α]_D²⁵ –2 ($c = 0.73$, CHCl₃); IR (NaCl, cm⁻¹) 3400, 3063, 3032, 2927, 2867, 1699, 1685, 1453, 1408, 1351, 1317, 1266, 1094, 1071, 1027; ¹H NMR (500 MHz, acetone-*d*₆) δ 3.31 (m, 1H, CH₂COPh), 3.55 (m, 2H, H6a + CH₂COPh), 3.71 (br s, 1H, H6b), 3.91 (m, 1H, H5), 4.21–4.28 (m, 3H, H3, H2 + OCH₂Ph), 4.39–4.45 (m, 3H, OCH₂Ph + H4), 4.54–4.65 (m, 4H, OCH₂Ph), 4.70–4.89 (m, 2H, OCH₂Ph + H1), 5.13 (br s, 2H, OCH₂Ph), 6.87–7.58 (m, 30H, CH_{Ar}'s); ¹³C NMR (125 MHz, acetone-*d*₆) δ 39.9 (br, CH₂COPh), 59.9–60.4 (C1), 66.5 (C4), 68.0 (OCH₂Ph), 72.4–72.6 (C6), 73.85, 73.93, 74.4, and 79.3 (OCH₂Ph), 82.3 (C3), 83.4–83.5 (C2), 127.5–131.2

(CH_{Ar}'s), 134.4–134.8 (CH_{Ar}'s), 138.7, 138.9, 139.3, 140.0, 140.2, and 140.7 (C_{qAr}), 157.6 (C=O), 199.9 (C=O); (+) MS (ESI) : $m/z = 794.0$ [M + NH₄]⁺; HRMS (ESI) calcd for C₅₀H₄₉NNaO₇: $m/z = 798.3407$ [M + Na]⁺; found : $m/z = 798.3410$.

Note: A sample of mesylated intermediate **4c** was purified by flash chromatography (PE/AcOEt 4:1, 70% yield). Cyclization of **4c** according to procedure **B** provided **5c** in 21% yield.

1(R)-2,3,5,6-Tetra-O-benzyl-N-benzyloxycarbonyl-1,4-dideoxy-1,4-imino-1-C-(2-oxocyclohexyl)-D-galactitol (5d). According to general procedure **B** applied to compound **3d** (649 mg, 0.841 mmol), mesylated intermediate **4d** was isolated (223.1 mg, 32%) as a colorless oil after purification by flash chromatography (PE/AcOEt 4:1). Cyclization of intermediate **4d** (184 mg, 0.228 mmol) according to procedure **B** afforded compound **5d** as a colorless oil (61.5 mg, 36%) after purification by chromatography (PE/AcOEt 4:1). Alternatively, starting from crude **3d**, compound **5d** was obtained in 36% yield after purification by flash chromatography, without purification of intermediate **4d**. Compound **5d**: [α]_D²⁵ +31 ($c = 0.94$, CHCl₃); IR (NaCl, cm⁻¹) 3063, 3031, 2935, 2862, 1703, 1453, 1402, 1351, 1307, 1213, 1093, 1027; ¹H NMR (250 MHz, DMSO-*d*₆, 80 °C) δ 1.11–1.65 (m, 4H), 1.67–2.31 (m, 4H), 2.69–2.81 (m, 1H, CHCO), 3.39–3.79 (m, 3H, 2H6 and H5), 4.02–4.17 (m, 2H, H3 and H2), 4.28–4.39 (m, 2H, H4 and OCH₂Ph), 4.39–4.56 (m, 6H, OCH₂Ph), 4.60–4.74 (m, 2H, OCH₂Ph and H1), 4.96–5.11 (m, 2H, OCH₂Ph), 7.16–7.35 (m, 25H, CH_{Ar}'s); ¹³C NMR (62.5 MHz, DMSO-*d*₆, 80 °C) δ 24.1, 27.8, 31.3, 41.5, and 50.3 (cyclohexyl CH₂ and CH), 58.6 (C1), 63.9 (C4), 66.0, 70.2, 70.6, 70.7, 71.9, 72.2, and 72.2 (OCH₂Ph, C6), 77.8 (C5), 82.1 (C3 or C2), 82.6 (C3 or C2), 127.0–127.8 (CH_{Ar}'s), 136.4, 137.5, 137.7, 137.9, and 138.2 (C_{qAr}), 156.3 (C=O), 210.5 (C=O); (+) MS (ESI) : $m/z = 754.5$ [M + H]⁺, 771.5 [M + NH₄]⁺.

1(R)-2,3,5,6-Tetra-O-benzyl-N-benzyloxycarbonyl-1,4-dideoxy-1,4-imino-1-C-(2-oxocyclopentyl)-D-galactitol (5e). According to general procedure **B** applied to compound **3e** (127 mg, 0.186 mmol), mesylated intermediate **4e** was obtained as a colorless oil (103 mg, 73%) after purification by flash chromatography (PE/AcOEt 7:3). Then compound **5e** was obtained from intermediate **4e** (83 mg, 0.1 mmol) as a colorless oil (18.3 mg, 25%, 67:33 mixture of diastereoisomers at cp-C2) after purification by flash chromatography (PE/AcOEt 4:1). IR (NaCl, cm⁻¹) 3019, 2401, 1727, 1720, 1701, 1454, 1406, 1353, 1216, 1093, 1071, 1028; ¹H NMR (500 MHz, acetone-*d*₆; d1 and d2 for diast. 1 and 2, respectively; cp for cyclopentyl) δ 1.52–2.15 (m, 6H, cp-CH₂), 2.32–2.40 (m, 1H, cp-CH), 3.52 (dd, 0.76H, $J \approx 11$ and 5.5 Hz, H6a, d1), 3.61 (dd, 0.34H, $J \approx 11$ and 5 Hz, H6a, d2), 3.72 (dd, 0.76H, $J \approx 11$ and 3 Hz, H6b, d1), 3.77 (dd, 0.34H, $J \approx 11$ and 3 Hz, H6b, d2), 4.02–4.06 (m, 1H, H5), 4.07 (d, 0.75H, $J \approx 6$ Hz, H2, d1), 4.16–4.27 (m, 1.35H, H3 and H2, d2), 4.36–4.52 (m, 5H, OCH₂Ph, H1 and H4), 4.54–4.66 (m, 4H, OCH₂Ph), 4.69–4.80 (m, 1H, OCH₂Ph), 4.97 (d, 0.7H, $J \approx 13$ Hz, OCH₂Ph), 5.00–5.11 (m, 1.3H, OCH₂-Ph), 7.24–7.44 (m, 25H, CH_{Ar}'s); ¹³C NMR (125 MHz, acetone-*d*₆; d1 and d2 for diast. 1 and 2, respectively; cp for cyclopentyl) δ 21.4 (d1, cp-C4), 21.5 (d2, cp-C4), 26.7 (d1, cp-C3), 28.4 (d2, cp-C3), 37.5 (d1, cp-C5), 38.9 (d2, cp-C5), 49.0–49.2 (cp-C2), 62.0–62.3 (C1), 66.5 (C4), 67.3, 67.4, 71.5, 71.6, 71.68, 71.74, 72.8, 73.0, 73.3, 73.66, and 73.73 (OCH₂Ph, C6), 79.1 (d2, C5), 79.2 (d1, C5), 80.3–80.5 (d1, C3), 80.58–80.63 (d2, C3), 84.2–84.6 (C2), 128.0–129.2 (CH_{Ar}'s), 138.0, 138.4, 138.89, 138.90, 139.3, 139.6, and 140.3 (C_{qAr}), 157.5 (C=O), cp-C1 hidden by solvent C=O signal; (+) MS (ESI) : $m/z = 762.0$ [M + Na]⁺; HRMS (ESI) calcd for C₄₇H₄₉NNaO₇: $m/z = 762.3407$ [M + Na]⁺; found : $m/z = 762.3416$.

1(R)-2,3,5,6-Tetra-O-benzyl-N-benzyloxycarbonyl-1-C-but-2-ynyl-1,4-dideoxy-1,4-imino-D-galactitol (5h). According to general procedure **B** applied to compound **3h** (30.1 mg, 0.037 mmol), compound **5h** was obtained as a colorless oil (16.2 mg, 63%) after purification by flash chromatography (PE/AcOEt 9:1) by way of mesylate **4h**. Compound **5h**: [α]_D²⁵ +1.5 ($c = 1.42$, CHCl₃); IR

(NaCl, cm⁻¹) 3424, 2921, 2861, 1695, 1454, 1403, 1353, 1095, 1070, 1027; ¹H NMR (400 MHz, CDCl₃) δ 1.70 (s, 3H, CH₃), 2.28–2.35 (m, 1H, CH_{2a}), 2.53–2.91 (m, 1H, CH_{2b}), 3.37–3.66 (m, 2H, H6), 3.85–3.97 (m, 1H, H5), 4.00–4.10 (m, 2H, H2, H3), 4.18–4.31 (m, 2H, H1, H4), 4.37–4.70 (m, 8H, OCH₂Ph), 5.04–5.20 (m, 2H, OCH₂Ph), 7.10–7.34 (m, 25H, CH_{Ar}'s); ¹³C NMR (100 MHz, CDCl₃) δ 3.7 (CH₃), 19.0–20.3 (CH₂), 60.5–61.9 (C1), 65.1 (C4), 67.2 (OCH₂Ph), 71.4 (OCH₂Ph), 71.5–71.8 (C6), 73.1, 73.2, and 73.4 (OCH₂Ph), 76.4 (C_{alkyne}-CH₃), 77.4 (C_{alkyne}-CH₂), 77.6 (C5), 80.8–81.0 (C2 or C3), 81.6–82.8 (C2 or C3), 127.6–128.6 (CH_{Ar}'s), 136.9, 137.9, 138.1, 138.4, and 138.8 (C_{qAr}), 156.8 (C=O); (+) MS (ESI) : $m/z = 712.5$ [M + H]⁺; HRMS (ESI) calcd for C₄₆H₄₇NNaO₆: $m/z = 732.3301$ [M + Na]⁺; found : $m/z = 732.3313$.

1(R)-1-C-Allenyl-2,3,5,6-tetra-O-benzyl-N-benzyloxycarbonyl-1,4-dideoxy-1,4-imino-D-galactitol (5i). According to general procedure **B** applied to compound **3i** (50 mg, 0.063 mmol), compound **5i** was obtained as a colorless oil (33.8 mg, 77%) after purification by flash chromatography (PE/AcOEt 9:1) by way of mesylate **4i**. Compound **5i**: [α]_D²⁵ +28 ($c = 0.61$, CHCl₃); IR (NaCl, cm⁻¹) 3415, 2929, 2867, 1957, 1702, 1497, 1454, 1405, 1353, 1267, 1097; ¹H NMR (400 MHz, CDCl₃) δ 3.54–3.75 (m, 2H, H6), 3.95–4.05 (m, 2H, H2 and H5), 4.07–4.20 (m, 2H, H3 and H4), 4.37–4.85 (m, 11H, OCH₂Ph, CH₂= and H1), 4.98–5.28 (m, 3H, OCH₂Ph and CH=), 7.18–7.35 (m, 25H, CH_{Ar}'s); ¹³C NMR (100 MHz, CDCl₃) δ 60.0 (C1), 63.5 (C4), 67.4, 72.1, 72.2, 73.3, 73.4 (OCH₂Ph, C6), 75.8–76.1 (CH₂=), 77.7 (C5), 81.9–82.1 (C3), 82.9–83.1 (C2), 88.2 (CH=), 127.6–128.5 (CH_{Ar}'s), 136.6, 137.8, 138.2, 138.5, and 138.7 (C_{qAr}), 156.1 (C=O), 209.3 (C=); (+) MS (ESI) : $m/z = 696.5$ [M + H]⁺, 714 [M + NH₄]⁺; HRMS (ESI) calcd for C₄₅H₄₅NNaO₆: $m/z = 718.31446$ [M + Na]⁺; found : $m/z = 718.3143$.

General Procedure C: Carbamate Deprotection. Compound **5a** or **5b** (0.056 mmol) and triethylamine (2 μ L) were dissolved in ethanol (0.5 mL). The flask was submitted to 3 freeze–pump–thaw cycles and palladium on charcoal (10% Pd on C, 0.1 equiv) was added under argon. The mixture was then placed under hydrogen. After a stirring time of 1.5–3 h at room temperature, the solids were removed by filtration on celite and washed with dichloromethane. Evaporation of the solvents provided a colorless oily product which was homogeneous by NMR.

1(R)-2,3,5,6-Tetra-O-benzyl-1,4-dideoxy-1,4-imino-1-C-propyl-D-galactitol (6a). This product was obtained from **5a** (39 mg, 0.056 mmol) as a colorless oil (31.2 mg, 99%). [α]_D²⁵ –33 ($c = 0.91$, CHCl₃); IR (NaCl, cm⁻¹) 3055, 2864, 1612, 1454, 1420, 1265, 1108; ¹H NMR (500 MHz, acetone-*d*₆) δ 0.91 (t, 3H, $J \approx 7.4$ Hz, CH₃), 1.29–1.46 (m, 2H, CH₂CH₃), 1.50–1.57 (m, 1H) and 1.59–1.67 (m, 1H) (CH₂CH₂CH₃), 2.97 (dt, 1H, $J \approx 7$ Hz, $J \approx 4$ Hz, H1), 3.14 (t, 1H, $J \approx 4$ Hz, H4), 3.72–3.74 (m, 2H, 2H6), 3.81 (d, 1H, $J \approx 4$ Hz, H2), 3.81–3.81 (m, 1H, H5), 3.92 (d, 1H, $J \approx 4.5$ Hz, H3), 4.18–4.25 (m, 0.5H, NH), 4.45–4.55 (m, 5H, OCH₂Ph), 4.60–4.66 (m, 2H, OCH₂Ph), 4.76 (d, 1H, $J \approx 11.7$ Hz, OCH₂-Ph), 7.21–7.37 (m, 20H, H_{Ar}); ¹³C NMR (125 MHz, acetone-*d*₆) δ 15.7 (CH₃), 22.1 (CH₂CH₃), 32.7 (CH₂CH₂CH₃), 63.9 (C1), 68.8 (C4), 72.3, 73.3, and 74.1 (CH₂OPh), 74.5 (C6), 74.7 (CH₂OPh), 79.7 (C5), 86.4 (C2), 86.9 (C3), 129.1–130.6 (CH_{Ar}'s), 130.6 (CH_{Ar}), 133.0 (CH_{Ar}), 140.76, 140.80, 140.8, and 141.0 (C_{qAr}); (+) MS (ESI) : $m/z = 566.5$ [M + H]⁺; HRMS (ESI) calcd for C₃₇H₄₃-NO₄: $m/z = 566.3270$ [M + H]⁺; found : $m/z = 566.3266$.

3,4,6,7-Tetra-O-benzyl-2,5-dideoxy-2,5-imino-D-glycero-1-glucoheptonitrile (6b). Deprotection of **5b** (12 mg, 0.018 mmol) according to general procedure **C** afforded **6b** as colorless oil (9.5 mg, 99%). [α]_D²⁵ –7 ($c = 0.81$, CHCl₃); IR (NaCl, cm⁻¹) 3019, 2927, C≡N not visible, 1602, 1455, 1215, 1096; ¹H NMR (500 MHz, CDCl₃) δ 3.09 (t, 1H, $J \approx 5.3$ Hz, H4), 3.54–3.61 (m, 2H, H6), 3.68 (dd, 1H, $J \approx 10$ Hz, $J \approx 5$ Hz, H5), 3.81 (dd, 1H, $J \approx 5$ Hz, $J \approx 3.2$ Hz, H3), 3.90 (d, 1H, $J \approx 5.2$ Hz, H1), 4.00 (dd, 1H,

$J \approx 3.1$ Hz, $J \approx 5.2$ Hz, H2), 4.21 (d, 1H, $J \approx 11.7$ Hz, OCH_2Ph), 4.33 (d, 1H, $J \approx 11.7$ Hz, OCH_2Ph), 4.40 (s, 2H, OCH_2Ph), 4.43 (d, 1H, $J \approx 11.7$ Hz, OCH_2Ph), 4.50 (d, 1H, $J \approx 11.8$ Hz, $\text{OCH}_2\text{-Ph}$), 4.57 (d, 1H, $J \approx 11.8$ Hz, OCH_2Ph), 4.66 (d, 1H, $J \approx 11.5$ Hz, OCH_2Ph), 7.10–7.31 (m, 20H, $\text{CH}_{\text{Ar's}}$); ^{13}C NMR (125 MHz, CDCl_3) δ 51.4 (C1), 65.0 (C4), 71.1 (C6), 72.3, 72.6, 73.1, and 73.6 (OCH_2Ph), 77.3 (C5), 83.4 (C2), 83.9 (C3), 118.0 (CN), 127.8–128.7 ($\text{CH}_{\text{Ar's}}$), 137.1, 137.7, 138.1, and 138.3 (C_{qAr}); (+) MS (ESI): $m/z = 550.0$ [$\text{M} + \text{H}$] $^+$; HRMS (ESI) calcd for $\text{C}_{35}\text{H}_{36}\text{N}_2\text{NaO}_4$: $m/z = 571.2573$ [$\text{M} + \text{Na}$] $^+$; found: $m/z = 571.2572$.

General Procedure D: O-Debenzylation. To a 0.01 M solution of benzylated compound (1 equiv) in dry CH_2Cl_2 was added at 0 °C a fresh 1 M solution of BCl_3 in hexane (11.5 equiv). The mixture was stirred overnight at 0 °C, and then the solids formed were dissolved by addition of MeOH (20 mL) and a few drops of water. The reaction mixture was then concentrated under reduced pressure. In some cases, it was found necessary to repeat the reaction with 3 equiv of BCl_3 overnight. In general, purification of the crude final product was performed by flash chromatography on C_{18} -reverse phase silica gel ($\text{H}_2\text{O}/\text{MeOH}$ 1:0 to 4:1), thus providing the debenzylated compound as its hydrochloride salt.

1(R)-1-C-Allyl-1,4-dideoxy-1,4-imino-D-galactitol (7a). According to general procedure D, compound **7a** was prepared from **5a** (134 mg, 0.19 mmol). MeOH and water (5 mL, 20:1) were added to the mixture at the end of the reaction and the solution was quickly filtered through ion-exchange resin (Dowex 1 \times 8, OH^- form) which was then washed with MeOH (2 \times 20 mL) and water (10 mL). Purification of the crude product by flash chromatography on silica gel ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 7:3 to 3:2) provided compound **7a** (12.7 mg, 33%) as a brown product. $[\alpha]_{\text{D}}^{25} -30$ ($c = 1.21$, MeOH); ^1H NMR (CD_3OD , 500 MHz) δ 2.48–2.54 (m, 1H) and 2.60–2.66 (m, 1H) (allyl- CH_2), 3.37 (dd, 1H, $J \approx 8$ Hz, $J \approx 2.5$ Hz, H4), 3.57 (td, 1H, $J \approx 3$ and 7.5 Hz, H1), 3.65 (dd, 1H, $J \approx 5$ and 11.5 Hz, H6a), 3.72 (dd, 1H, $J \approx 4.5$ and 11.5 Hz, H6b), 3.85–3.90 (m, 1H, H5), 3.98 (dd, 1H, $J \approx 1$ and 3 Hz, H2), 4.08 (dd, 1H, $J \approx 1$ and 2.5 Hz, H3), 5.18 (dd, 1H, $J \approx 1.5$ and 10 Hz, $=\text{CH}_a\text{H}_b$), 5.27 (dd, 1H, $J \approx 1.5$ and 17 Hz, $=\text{CH}_a\text{H}_b$), 5.89 (m, 1H, $=\text{CH}$); ^{13}C NMR (CD_3OD , 125 MHz) δ 31.5 ($\text{CH}_2\text{CH}=\text{}$), 63.1 (C1), 64.8 (C6), 70.3 (C4), 71.7 (C5), 77.0 (C2), 78.8 (C3), 119.0 ($=\text{CH}_2$), 134.5 ($=\text{CH}$); (+) MS (ESI): $m/z = 204.0$ [$\text{M} + \text{H}$] $^+$; HRMS (ESI) calcd for $\text{C}_9\text{H}_{18}\text{NO}_4$: $m/z = 204.1236$ [$\text{M} + \text{H}$] $^+$; found: $m/z = 204.1239$.

1(R)-2,3,5,6-Tetra-O-benzyl-N-benzyloxycarbonyl-1,4-dideoxy-1-C-(3-diethylphosphono-2-propen-1-yl)-1,4-imino-D-galactitol (9). To a solution of **5a** (172.4 mg, 0.248 mmol) in dry CH_2Cl_2 (4.1 mL) was added dropwise commercial diethyl vinylphosphonate (76.8 μL , 0.495 mmol, 2 equiv). The flask was submitted to 3 freeze–pump–thaw cycles and placed under Ar. Then Hoveyda-Grubbs catalyst (7.8 mg, 5%) was added and the 3 freeze–pump–thaw–argon cycles were repeated. The reaction mixture was stirred for 12 h at 40 °C, then another addition of Hoveyda-Grubbs catalyst (7.8 mg, 5%) was made. After an additional stirring time of 12 h at 40 °C, the reaction mixture was concentrated under reduced pressure. Excess diethyl vinylphosphonate was removed from the brown residue by coevaporation with toluene. Purification by flash chromatography ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 99:1) afforded compound **9** (154.9 mg, 75%) as a brown oily product. IR (NaCl, cm^{-1}) 3063, 3031, 2981, 2867, 1701, 1454, 1402, 1353, 1267, 1247, 1216, 1099, 1059, 1027; ^1H NMR (400 MHz, $\text{DMSO}-d_6$, 80 °C; $J_{\text{H,H}}$ coupling constants were measured on a ^{31}P -decoupled spectrum) δ 1.22 (t, 6H, $J \approx 7$ Hz, CH_3), 2.48–2.53 (m, 1H) and 2.58–2.66 (m, 1H) ($\text{CH}_2\text{CH}=\text{CH}$), 3.57 (dd, 1H, $J \approx 5.9$ Hz, $J \approx 10.9$ Hz, H6a), 3.70 (dd, 1H, $J \approx 2.8$ Hz, $J \approx 10.9$ Hz, H6b), 3.87 (m, 1H, H5), 3.92 (quint, 4H, $J \approx 7.1$ Hz, CH_2CH_3), 4.09–4.13 (m, 1H, H2), 4.16–4.19 (m, 2H, H3, H4), 4.29 (q, 1H, $J \approx 7.2$ Hz, H1), 4.43–4.68 (m, 8H, OCH_2Ph), 5.09 (broad s, 2H, OCH_2Ph), 5.61–5.70 (dd, 1H, $J \approx 17$ Hz, $J_{\text{H-P}} \approx 21.3$ Hz, $\text{PCH}=\text{CH}$), 6.58–6.71 (ddt, 1H, $J \approx 6.6$ Hz, $J \approx 17$ Hz, $J_{\text{H-P}} \approx 21.8$ Hz, $\text{PCH}=\text{CH}$), 7.28–7.37 (m,

25H, $\text{CH}_{\text{Ar's}}$); ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$, 80 °C) δ 16.5 (d, CH_3), 34.2 (d, $J \approx 21.6$ Hz, $\text{CH}_2\text{CH}=\text{CHP}$), 59.8 (C1), 61.4 (d, $\text{CH}_3\text{CH}_2\text{O}$), 64.4 (C2), 66.9 (OCH_2Ph), 71.4 (OCH_2Ph), 71.8 (C6), 72.4 (OCH_2Ph), 72.9 (OCH_2Ph), 73.1 (OCH_2Ph), 78.2 (C5), 81.9 (C3 or C4), 82.5 (C3 or C4), 118.8 ($\text{PCH}=\text{CH}_2$, $J_{\text{C,P}} \approx 186$ Hz, measured in CDCl_3), 127.7–128.7 ($\text{CH}_{\text{Ar's}}$), 137.8–138.2 (C_{qAr}), 150.0 ($\text{CH}_2\text{CH}=\text{CH}$), 156.4 ($\text{C}=\text{O}$); ^{31}P NMR (202 MHz, CDCl_3) δ 18.25; (+) MS (ESI): $m/z = 835$ [$\text{M} + \text{H}$] $^+$, 856 [$\text{M} + \text{Na}$] $^+$; HRMS (ESI) calcd for $\text{C}_{49}\text{H}_{56}\text{NNaO}_9\text{P}$: $m/z = 856.3590$ [$\text{M} + \text{Na}$] $^+$; found: $m/z = 856.3583$. Anal. Calcd. for $\text{C}_{49}\text{H}_{56}\text{NO}_9\text{P}$: C, 70.57; H, 6.77; N, 1.68. Found: C, 70.15; H, 6.70; N, 1.71.

1,4-Bis-[1(R)-2,3,5,6-tetra-O-benzyl-N-benzyloxycarbonyl-1,4-dideoxy-1,4-imino-D-galactitol-1-yl]-2-butene (10). A solution of **5a** (209 mg, 0.3 mmol) in dry CH_2Cl_2 (3.5 mL) was submitted to 3 freeze–pump–thaw cycles and placed under Ar. Then Hoveyda-Grubbs catalyst (9.4 mg, 5%) was added and the 3 freeze–pump–thaw–argon cycles were repeated. The green reaction medium was stirred overnight at 40 °C and then the brown mixture was concentrated under reduced pressure. Purification by flash chromatography (PE/AcOEt 85:15) provided compound **10** (108.1 mg, 53%) as a green oily product. IR (NaCl, cm^{-1}) 3442, 3052, 3031, 1695, 1454, 1403, 1265, 1096, 1069; ^1H NMR (400 MHz, CDCl_3) δ 2.17–2.31 (m, 2H, $\text{CH}_2\text{CH}=\text{}$), 2.37–2.77 (m, 2H, $\text{CH}_2\text{CH}=\text{}$), 3.39–3.68 (m, 4H, H6), 3.84–3.96 (m, 4H, H5, H2), 3.99–4.14 (m, 4H, H1, H3), 4.20 (dd, 2H, $J \approx 2.4$ and 6.8 Hz, H4), 4.28–4.56 (m, 14H, OCH_2Ph), 4.58–4.70 (m, 2H, OCH_2Ph), 4.97–5.13 (m, 4H, OCH_2Ph), 5.39–5.50 (m, 2H, $\text{CH}=\text{}$), 7.16–7.29 (m, 50H, $\text{CH}_{\text{Ar's}}$); ^{13}C NMR (62.5 MHz, CDCl_3) δ 27.7 ($\text{CH}_2\text{CH}=\text{}$), 32.4, ($\text{CH}_2\text{CH}=\text{}$), 60.6–61.6 (C1), 64.1 (C4), 64.4 (C4), 67.06 (C6), 67.10 (C6), 71.5, 71.6, 72.0–72.1, 72.4, 73.15, 73.21, and 73.3 (OCH_2Ph), 77.4 (C5), 77.9 (C5), 80.8 (C3), 81.12–81.15 (C3), 82.3–82.8 (C2), 127.5–128.5 ($\text{CH}_{\text{Ar's}}$), 129.1 ($\text{CH}_2\text{CH}=\text{}$), 129.2 ($\text{CH}_2\text{CH}=\text{}$), 136.82, 136.84, 137.8, 138.1, 138.2, 138.4, 138.7, and 138.8 (C_{qAr}), 156.8 ($\text{C}=\text{O}$); (+) MS (ESI): $m/z = 1385.0$ [$\text{M} + \text{NH}_4$] $^+$; HRMS (ESI) calcd for $\text{C}_{88}\text{H}_{90}\text{N}_2\text{O}_{12}\text{Na}$: $m/z = 1389.63915$ [$\text{M} + \text{Na}$] $^+$; found: $m/z = 1389.6389$. Anal. Calcd. for $\text{C}_{88}\text{H}_{90}\text{N}_2\text{O}_{12}$: C, 77.28; H, 6.63; N, 2.05. Found: C, 77.06; H, 6.65; N, 1.98.

1(R)-1,4-Dideoxy-1-C-(3-diethylphosphono-2-propen-1-yl)-1,4-imino-D-galactitol (11). Compound **11** was obtained according to procedure D, starting from compound **9** (220 mg, 0.263 mmol). Purification by C_{18} -reverse phase silica gel chromatography ($\text{H}_2\text{O}/\text{MeOH}$ 1:0 to 9:1) afforded the hydrochloride salt of **11** (98.8 mg, 74%) as a colorless foamy product. $[\alpha]_{\text{D}}^{25} -20$ ($c = 1.46$, MeOH); ^1H NMR (500 MHz, CD_3OD) δ 1.33 (t, 6H, $J \approx 7$ Hz, CH_3), 2.74–2.80 (m, 1H) and 2.85–2.91 (m, 1H) ($\text{CH}_2\text{CH}=\text{CH}$), 3.44–3.48 (m, 1H, H4), 3.66 (dd, 1H, $J \approx 4.5$ Hz, $J \approx 11.5$ Hz, H6a), 3.71–3.77 (m, 2H, H1 and H6b), 3.86–3.90 (m, 1H, H5), 4.01 (d, 1H, $J \approx 2.5$ Hz, H2), 4.06–4.13 (m, 5H, H3 and CH_2CH_3), 6.02 (dd, 1H, $J \approx 17$ Hz, $J_{\text{H-P}} \approx 20.8$ Hz, $\text{PCH}=\text{CH}$), 6.75 (ddt, 1H, $J \approx 7$ Hz, $J \approx 17$ Hz, $J_{\text{H-P}} \approx 21.5$ Hz, $\text{PCH}=\text{CH}$); ^{13}C NMR (125 MHz, CD_3OD) δ 16.6 (CH_3), 16.7 (CH_3), 31.4 (d, $J \approx 23.7$ Hz, $\text{CH}_2\text{-CH}=\text{CHP}$), 62.1 (C1), 63.5 ($\text{CH}_3\text{CH}_2\text{O}$), 63.6 ($\text{CH}_3\text{CH}_2\text{O}$), 64.7 (C6), 70.9 (C4), 71.5 (C5), 76.9 (C2), 78.1 (C3), 121.8 (d, $J \approx 186.5$ Hz, $\text{PCH}=\text{CH}_2$), 148.9 (d, $J \approx 5.3$ Hz, $\text{CH}_2\text{CH}=\text{CH}$); ^{31}P NMR (202 MHz, CD_3OD) δ 17.84 (s); (+) MS (ESI): $m/z = 340.5$ [$\text{M} + \text{H}$] $^+$, 362.5 [$\text{M} + \text{Na}$] $^+$; HRMS (ESI) calcd for $\text{C}_{13}\text{H}_{26}\text{-NNaO}_7\text{P}$: $m/z = 362.13446$ [$\text{M} + \text{Na}$] $^+$; found: $m/z = 362.1340$. Anal. Calcd. for $\text{C}_{13}\text{H}_{27}\text{ClNO}_7\text{P} + 2\text{H}_2\text{O}$: C, 39.65; H, 7.42; N, 3.56; Cl, 9.00. Found: C, 39.75; H, 6.94; N, 3.4; Cl, 9.66.

General Procedure E: Preparation of Free Phosphonic Acids. To a 0.1 M solution of ethyl phosphonate (**11** or **19**) (1 equiv) in dry MeCN was added BrSiMe_3 (10 eq) at room temperature. The mixture was stirred for the indicated time. The mixture was then diluted with MeOH (15 mL) and concentrated under reduced pressure. The dilution and evaporation process was repeated twice. The residue was then taken up with water and the mixture extracted with CH_2Cl_2 to remove traces of less polar organic compounds. The aqueous phase was concentrated under reduced pressure.

Chromatography of the crude product on C₁₈-reversed phase silica gel (H₂O/MeOH 1:0 to 4:1) provided free phosphonic acids.

(1R)-1,4-Dideoxy-1-C-(3-phosphono-2-propen-1-yl)-1,4-imino-D-galactitol (12). Compound **12** was obtained as a brownish foam (10.7 mg, 59%) from **11** (20 mg, 0.05 mmol) according to general procedure **E** (reaction time: 64 h). ¹H NMR (400 MHz, CD₃OD) δ 2.70–2.77 (m, 1H) and 2.81–2.89 (m, 1H) (CH₂CH=CH), 3.47 (dd, 1H, *J* ≈ 2.4 and 8.8 Hz, H4), 3.67 (dd, 1H, *J* ≈ 8 Hz, *J* ≈ 11.6 Hz, H6a), 3.70–3.77 (m, 2H, H1 and H6b), 3.88–3.92 (m, 1H, H5), 4.02–4.07 (m, 1H, H2), 4.13 (m, 1H, *J* ≈ 2.4 Hz, H3), 6.01–6.11 (m, 1H, PCH=CH), 6.54–6.70 (m, 1H, PCH=CH); ¹³C NMR (100 MHz, CD₃OD) δ 31.1 (d, *J* ≈ 23 Hz, CH₂CH=CHP), 62.3 (C1), 64.6 (C6), 70.7 (C4), 71.5 (C5), 76.7 (C2), 78.1 (C3), 117.5 (PCH=CH₂), 144.7 (CH₂CH=CH); ³¹P NMR (162 MHz, CD₃OD) δ 14.54 (s); HRMS (ESI) calcd for C₁₃H₂₆NNaO₇P: *m/z* = 362.13446 [M + Na]⁺; found: *m/z* = 362.1340.

1,4-Bis-[1(R)-1,4-dideoxy-1,4-imino-D-galactit-1-yl]-2-butene (13). Compound **13** was prepared by the debenzoylation procedure **D**, starting from compound **10** (196 mg, 0.143 mmol). Compound **13** was obtained as a colorless foam, as its hydrochloride salt (62 mg, 96%) and as a mixture of *Z/E* isomers (1:2). ¹H NMR (250 MHz, CD₃OD) δ 2.48–2.83 (m, 4H, CH₂CH=), 3.40–3.53 (m, 2H, H4), 3.57–3.81 (m, 6H, H1, 2H6), 3.86–3.94 (m, 2H, H5), 3.98–4.07 (m, 2H, H2), 4.07–4.18 (m, 2H, H3), 5.63–5.69 (m, 0.7H, CH=, *Z*-isomer), 5.71–5.80 (m, 1.3H, CH=, *E*-isomer); ¹³C NMR (62.5 MHz, CD₃OD) δ 24.9 (CH₂CH=, *Z*), 29.9 (CH₂CH=, *E*), 63.25 (C1, *Z*), 63.30 (C1, *E*), 64.6 (C6), 70.56 (C4, *E*), 70.60 (C4, *Z*), 71.45 (C5, *Z*), 71.49 (C5, *E*), 76.6 (C2, *Z*), 76.7 (C2, *E*), 78.2 (C3, *E*), 78.3 (C3, *Z*), 128.5 (CH=, *Z*), 130.0 (CH=, *E*); (+) MS (ESI): *m/z* = 379.5 [M + H]⁺; HRMS (ESI) calcd for C₁₆H₃₀N₂NaO₈: *m/z* = 401.18999 [M + Na]⁺, found: *m/z* = 401.1898.

Ethyl (2',3'-O-isopropylideneuridin-5'-yl) vinylphosphonate (17). To a solution of diethyl vinylphosphonate (377 μL, 2.44 mmol) in dry CH₂Cl₂ (10 mL) was added oxalyl chloride (230 μL, 2.68 mmol, 1.1 equiv) under Ar. After stirring overnight at room temperature, the mixture was stirred at 35 °C for 24 h and then concentrated under reduced pressure. Ethyl vinylphosphonyl chloride was thus obtained with a conversion of 94% (¹H NMR integration) as a colorless oily product. This reagent was used without further purification because it was air and moisture sensitive. To a solution of ethyl vinylphosphonyl chloride (340 mg, 2.2 mmol) in dry CH₂Cl₂ (11 mL) under Ar were added 2',3'-O-isopropylideneuridine (940 mg, 3.30 mmol, 1.5 equiv) and Et₃N (618 μL, 4.39 mmol, 2 equiv). The purple mixture was stirred for 36 h at 35 °C, then the reaction was quenched by adding water (2 mL). Compound **17** was extracted with CH₂Cl₂ (3×), the organic layers were combined and dried over Na₂SO₄, and the solvent was evaporated under reduced pressure. The pink foam thus obtained was purified by flash chromatography (CH₂Cl₂/MeOH 99:1 to 96:4). Compound **17** was obtained as a white foam (540 mg, 61% from commercial diethyl vinylphosphonate) and as a 1:1 mixture of *P*-stereoisomers (d1 and d2). IR (NaCl, cm⁻¹) 3466, 3169, 3057, 2989, 2940, 1694, 1633, 1457, 1383, 1271, 1235, 1159, 1067, 1019; ¹H NMR (250 MHz, CDCl₃) δ 1.21–1.27 (m, 6H, CH₃ isopropylidene + CH₃CH₂O), 1.48 (s, 3H, CH₃ isopropylidene), 3.98–4.11 (dq, 2H, *J* ≈ 7.5 Hz, CH₃CH₂O d1 and d2), 4.24–4.29 (m, 2H, H5'), 4.32–4.40 (m, 1H, H4'), 4.77–4.80 (m, 1H, H3'), 4.86 (dd, 1H, *J* ≈ 6.25 Hz, H2' d1 and d2), 5.64 (d, 1H, *J* ≈ 8 Hz, H5), 5.73 (dd, 1H, *J* ≈ 4.75 Hz, H1' d1 and d2), 5.87–6.39 (m, 3H, CH₂=CHP + CH₂=CHP d1 and d2), 7.39 (t, 1H, *J* ≈ 8.25 Hz, H6 d1 and d2), 10.44 (broad s, 1H, NH); ¹³C NMR (62.5 MHz, CDCl₃) δ 16.0, 16.1 (CH₃, d1 and d2), 26.0 and 26.9 (CH₃, isopropylidene), 62.2, 62.3 (CH₃CH₂O d1 and d2), 64.9–65.0 (m, C5' d1 and d2), 80.5, 80.6 (C3' d1 and d2), 84.2, 84.3 (C2' d1 and d2), 85.3 (d, *J*_{C-P} ≈ 11.3 Hz, C4' d1), 85.5 (d, *J*_{C-P} ≈ 11.3 Hz, C4' d2), 93.4, 93.7 (C1' d1 and d2), 102.2, 102.3 (C5, d1 and d2), 114.15, 114.18 (Cq isopropylidene, d1 and d2), 124.8 (d, *J*_{C-P} ≈ 183 Hz, PCH=CH₂ d1), 124.9 (d, *J*_{C-P} ≈ 183 Hz, PCH=CH₂

d2), 136.4 (PCH=CH₂), 141.8, 141.9 (C6, d1 and d2), 150.13, 150.15 (C2, d1 and d2), 163.7 (C4); ³¹P NMR (202 MHz, CDCl₃) δ 17.83 (s), 18.0 (s); (+) MS (ESI): *m/z* = 420.0 [M + NH₄]⁺; HRMS (ESI): calcd for C₁₆H₂₃N₂NaO₈P: *m/z* = 425.1090 [M + Na]⁺; found: *m/z* = 425.1092.

1(R)-2,3,5,6-Tetra-*O*-benzyl-*N*-benzyloxycarbonyl-1,4-dideoxy-1-C-[3-[(ethyl)(2',3'-O-isopropylideneuridin-5'-yl)phosphono]-2-propen-1-yl]-1,4-imino-D-galactitol (18). To a solution of compound **5a** (115 mg, 0.165 mmol) in dry CH₂Cl₂ (2.75 mL) was added compound **17** (133 mg, 0.33 mmol, 2 equiv). The flask was then submitted to 3 freeze–pump–thaw cycles and placed under Ar. Then Hoveyda-Grubbs catalyst (5.2 mg, 5%) was added and the 3 freeze–pump–thaw–argon cycles were repeated. The reaction medium was stirred for 24 h at 40 °C, then a second addition of Hoveyda-Grubbs catalyst (5.2 mg, 5%) was performed. After 20 h stirring time at 40 °C, the reaction mixture was concentrated under reduced pressure. The brownish residue was dried by coevaporation with toluene. Purification by flash chromatography (CH₂Cl₂/MeOH 99:1 to 98:2) provided compound **18** (107.5 mg, 61%) as a brown oily product (1:1 mixture of *P*-stereoisomers, d1 and d2). IR (NaCl, cm⁻¹) 3032, 2988, 2935, 2249, 1695, 1455, 1266, 1215, 1158, 1095, 1068, 1028; ¹H NMR (500 MHz, DMSO-*d*₆, 80 °C) δ 1.22 (t, 3H, *J* ≈ 6.3 Hz, OCH₂CH₃, d1), 1.24 (t, 3H, *J* ≈ 6.3 Hz, OCH₂CH₃, d2), 1.34 (s, 3H) and 1.54 (s, 3H, CH₃ isopropylidene), 2.48–2.53 (m, 1H, CH_aCH=CHP), 2.61–2.68 (m, 1H, CH_bCH=CHP), 3.58 (dd, *J* ≈ 5.8 Hz, *J* ≈ 10.9 Hz, 1H, H6a), 3.69 (dd, *J* ≈ 3.24 Hz, *J* ≈ 10.9 Hz, 1H, H6b), 3.88–3.94 (m, 1H, H5), 3.93–4.00 (m, 2H, OCH₂CH₃), 4.08–4.13 (m, 2H, H3 + H5'), 4.18 (m, 2H, H2 + H4), 4.22 (q, 1H, *J* ≈ 5.2 Hz, H4'), 4.27–4.33 (m, 1H, H1), 4.46 (d, 1H, *J* ≈ 5.1 Hz, OCH₂Ph), 4.50–4.59 (m, 6H, OCH₂Ph), 4.66 (d, 1H, *J* ≈ 11.6 Hz, OCH₂Ph), 4.82 (dd, 1H, *J* ≈ 4 Hz, *J* ≈ 6.4 Hz, H3'), 5.02 (m, 1H, H2'), 5.10 (broad s, 2H, OCH₂Ph), 5.61 (d, 1H, *J* ≈ 8 Hz, H5''), 5.66–5.74 (m, 1H, CH=CHP), 5.86–5.87 (m, 1H, H1'), 6.62–6.76 (m, 1H, CH=CHP), 7.27–7.39 (m, 25H, CH_{Ar}s), 7.64 (dd, 1H, *J* ≈ 8 Hz, H6''); ¹³C NMR (125 MHz, DMSO-*d*₆, 80 °C) δ 16.28 (CH₃), 16.32 (CH₃), 25.6 and 27.3 (2 CH₃ isopropylidene), 34.1 (d, *J*_{C-P} ≈ 22.1 Hz, CH₂CH=CHP), 59.66, 59.71 (C-1, d1 and d2), 61.66, 61.71 (OCH₂CH₃, d1 and d2), 64.26, 64.31 (C2, d1 and d2), 64.96, 65.00 (C5', d1 and d2), 66.8 (OCH₂-Ph), 71.2, 71.5, 72.3, 72.7, 72.9 (C6 + 4 OCH₂Ph), 78.0 (C5), 81.1 (C3'), 81.8 (C4), 82.3 (C-3), 83.9 (C2'), 85.2 (m, C4'), 92.65, 92.68 (C-1', d1 and d2), 102.3 (C5''), 113.8 (Cq isopropylidene), 118.5 (d, *J*_{C-P} ≈ 185.6 Hz, CH=CHP d1), 118.7 (d, *J*_{C-P} ≈ 185.6 Hz, CH=CHP d2), 127.5–128.5 (CH_{Ar}s), 137.1, 138.0, 138.5, 138.7, and 138.9 (Cq_{Ar}), 142.4 (C-6''), 150.6 (d, *J*_{C-P} ≈ 21.6 Hz, CH=CHP), 156.2 (C=O), 163.2 (C=O); ³¹P NMR (202 MHz, CDCl₃) δ 17.94 (s), 18.01 (s); (+) MS (ESI): *m/z* = 1095.5 [M + Na]⁺; HRMS (ESI) calcd for C₅₉H₆₆N₃NaO₁₄P: *m/z* = 1094.4180 [M + Na]⁺; found: *m/z* = 1094.4175. Anal. Calcd. for C₅₉H₆₆N₃O₁₄P: C, 66.09; H, 6.20; N, 3.92. Found: C, 66.07; H, 6.20; N, 3.79.

1(R)-1,4-Dideoxy-1-C-[3-[(ethyl)(uridin-5'-yl)phosphono]-2-propen-1-yl]-1,4-imino-D-galactitol (19). To a solution of **18** (79 mg, 0.073 mmol) in dry CH₂Cl₂ (2.5 mL) was added BCl₃ (862 μL, 735 mmol, 100 equiv) at 0 °C under argon. The reaction mixture was stirred overnight at 0 °C, then it was diluted by the addition of CH₂Cl₂ (2.5 mL) and the solids formed during the reaction were dissolved by the addition of MeOH (15 mL) and a few drops of water. The reaction mixture was then concentrated under reduced pressure. MeOH and water (5 mL, 20:1) were added and the solution was quickly filtered through ion-exchange resin (Dowex 1 × 8, OH⁻ form) which was then washed with MeOH (2 × 20 mL) and water (10 mL). Purification of the crude product by flash chromatography on C₁₈-reversed phase silica gel (H₂O/*i*-PrOH 1:0 to 7:3) provided compound **19** (20.2 mg, 52%) as a white foam (1:1 mixture of *P*-stereoisomers). ¹H NMR (500 MHz, CD₃OD, d1 and d2 for *P*-epimers) δ 1.35 (t, 3H, *J* ≈ 6.3 Hz, OCH₂CH₃, d1), 1.36 (t, 3H, *J* ≈ 6.3 Hz, OCH₂CH₃, d2), 2.70–2.78 (m, 1H) and 2.83–2.88 (m, 1H)(CH₂CH=CHP), 3.40 (m, 1H, H4), 3.63–

3.74 (m, 3H, H1, 2H6), 3.85–3.88 (m, 1H, H5), 3.99–4.00 (m, 1H, H2), 4.09 (brs, 1H, H3), 4.12–4.18 (m, 4H, OCH₂CH₃ + H2'), 4.19–4.21 (m, 1H, H4'), 4.22–4.26 (m, 1H, H5'a), 4.30–4.34 (m, 1H, H5'b), 5.71 (d, 1H, $J \approx 8$ Hz, H5''), 5.83 (dd, 1H, $J \approx 3.9$ Hz, $J \approx 6.1$ Hz, H1'), 6.01–6.09 (m, 1H, CH=CHP), 6.68–6.77 (m, 1H, CH=CHP), 7.72 (dd, 1H, $J \approx 8$ Hz, H6'' d1 and d2); ¹³C NMR (125 MHz, CD₃OD) δ 16.65, 16.70 (CH₃, d1 and d2), 32.0 (d, $J_{C-P} \sim 22.1$ Hz, CH₂CH=CHP), 61.8 (C1), 63.9, 64.0 (OCH₂CH₃, d1 and d2), 64.8 (C2), 66.2–66.3 (m, C5'), 70.5 (C4), 70.9 (C3'), 71.7 (C5), 74.99, 75.02 (C2', d1 and d2), 77.1 (C3), 78.6, 78.7 (C2, d1 and d2), 83.55, 83.60 (C4', d1 and d2), 91.8, 91.9 (C1', d1 and d2), 102.9 (C5''), 120.7 (d, $J_{C-P} \sim 187.3$ Hz, CH=CHP), 120.8 (d, $J_{C-P} \sim 187.1$ Hz, CH=CHP), 142.49, 142.54 (C-6'', d1 and d2), 150.5 (m, CH=CHP), 152.2 (C=O), 166.0 (C=O); ³¹P NMR (202 MHz, CD₃OD) δ 18.35 (s), 18.53 (s); (+) MS (ESI): $m/z = 538.5$ [M + H]⁺; HRMS (ESI) calcd for C₂₀H₃₃N₃O₁₂P: $m/z = 538.1802$ [M + H]⁺; found: $m/z = 538.1800$. Calcd for C₂₀H₃₂N₃NaO₁₂P: $m/z = 560.1621$ [M + Na]⁺; found: $m/z = 560.1622$.

Procedure for 19·HCl. Debenzylation of phosphonate **18** (149 mg, 0.139 mmol) according to general procedure **D** afforded compound **19** as its hydrochloride salt (62.6 mg, 78%). Anal. Calcd. for C₂₀H₃₃ClN₃O₁₂P+2H₂O: C, 39.38; H, 6.11; N, 6.89; Cl, 5.81. Found: C, 39.27; H, 6.29; N, 6.53; Cl, 5.80.

1(R)-1,4-Dideoxy-1-C-{3-[(uridin-5'-yl)phosphono]-2-propen-1-yl}-1,4-imino-D-galactitol (20). Compound **20** (8 mg, 36%) was obtained as a white foam from **19** (21.9 mg, 0.038 mmol) according to general procedure **E** (stirring time: 3 d). $[\alpha]_D^{25} -2$ ($c = 0.49$, MeOH); ¹H NMR (400 MHz, CD₃OD) δ 2.58–2.80 (m, 2H, CH₂-

CH=CHP), 3.42 (dd, 1H, $J \approx 2.4$ Hz, $J \approx 8.4$ Hz, H4), 3.62–3.77 (m, 3H, H1 and 2H6), 3.85–3.90 (m, 1H, H5), 3.98–4.12 (m, 5H), 4.19–4.23 (m, 2H), 5.79 (d, 1H, $J \approx 8$ Hz, H5''), 5.89–6.01 (m, 2H, H1 and CH=CHP), 6.36–6.50 (m, 1H, CH=CHP), 8.01 (d, 1H, $J \approx 8$ Hz, H6''); ¹³C NMR (100 MHz, CD₃OD) δ 31.4 (d, $J_{C-P} \sim 21.6$ Hz, CH₂CH=CHP), 62.8 (C1), 64.5 (d, $J_{C-P} \sim 4.6$ Hz, C5'), 64.7 (m, C6), 70.7 (C4), 71.5 (C3' or C2'), 71.7 (C5), 75.6 (C2' or C3'), 77.0 (C2), 78.3 (C3), 85.1 (d, $J_{C-P} \sim 8.2$ Hz, C4'), 90.0 (C1'), 103.0 (C5''), 128.4 (d, $J_{C-P} \sim 175$ Hz, CH=CHP), 142.1 (CH=CHP), 142.6 (C6''), 152.6 (C=O), 166.2 (C=O); ³¹P NMR (162 MHz, CD₃OD) δ 12.05 (s); (+) MS (ESI): $m/z = 532.5$ [M + Na]⁺; HRMS (ESI) calcd for C₁₈H₂₈N₃NaO₁₂P: $m/z = 532.13083$ [M + Na]⁺; found: $m/z = 538.1306$.

Inhibition Assay. Inhibition assays of UGM by the new compounds were performed as previously described.^{15b}

Acknowledgment. Funding of this research by CNRS and by the French Ministry of Research and Higher Education is gratefully acknowledged.

Supporting Information Available: Experimental procedures and characterization data for compounds **2**, **2'**, **3f**, **3f'**, **3g**, **8**, **14**, **15**, **16**, and copies of NMR spectra of all new compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

JO8001134